



## Stress and cancer: mechanisms, significance and future directions

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**Abstract** | The notion that stress and cancer are interlinked has dominated lay discourse for decades. More recent animal studies indicate that stress can substantially facilitate cancer progression through modulating most hallmarks of cancer, and molecular and systemic mechanisms mediating these effects have been elucidated. However, available clinical evidence for such deleterious effects is inconsistent, as epidemiological and stress-reducing clinical interventions have yielded mixed effects on cancer mortality. In this Review, we describe and discuss specific mediating mechanisms identified by preclinical research, and parallel clinical findings. We explain the discrepancy between preclinical and clinical outcomes, through pointing to experimental strengths leveraged by animal studies and through discussing methodological and conceptual obstacles that prevent clinical studies from reflecting the impacts of stress. We suggest approaches to circumvent such obstacles, based on targeting critical phases of cancer progression that are more likely to be stress-sensitive; pharmacologically limiting adrenergic-inflammatory responses triggered by medical procedures; and focusing on more vulnerable populations, employing personalized pharmacological and psychosocial approaches. Recent clinical trials support our hypothesis that psychological and/or pharmacological inhibition of excess adrenergic and/or inflammatory stress signalling, especially alongside cancer treatments, could save lives.

For decades, stress has been suggested to affect cancer incidence and cancer progression<sup>1,2</sup>. However, both epidemiological studies and clinical trials have yielded mixed results, or indicated small or clinically insignificant effects of stress on cancer progression. Consequently, current medical routines do not include measures to prevent stress responses as a means to improve cancer survival. Within the medical community, this may reflect a disbelief that stress is a significant biological factor underlying cancer aetiology and progression.

By contrast, in recent years, animal studies have provided solid evidence that stress can facilitate growth and metastasis of many types of cancer. Most importantly, numerous endocrine, cellular and molecular mechanisms underlying these effects have been identified. For example, animal models have shown that stress factors can promote most established hallmarks of cancer<sup>2</sup>, and that stress responses can facilitate cancer growth and metastasis via directly affecting molecular characteristics of the malignant tissue<sup>3–5</sup>, its microenvironment<sup>6</sup>, antitumour immune activity<sup>4,7–9</sup> and other indirect modulators of cancer progression<sup>10,11</sup>. In patients with cancer, stress has been shown to activate many of these

processes<sup>8,10–13</sup>, supporting the clinical significance of these findings.

We suggest that the discrepancy between preclinical studies and clinical or epidemiological studies stems from two sources. First, preclinical studies can synchronize stress or stress-reducing interventions with critical periods along cancer progression that are highly susceptible to the impacts of stress. Second, conceptual and methodological difficulties in conducting clinical studies may obscure the impact of stress on cancer progression.

In this Review, we describe and discuss stress and stress responses at the organism level and in the context of cancer. We further explain mechanisms via which stress can facilitate cancer initiation, impair cancer treatments and promote cancer growth and metastasis, based on animal studies and on parallel human correlative or causative studies. We also review epidemiological studies and clinical trials in patients with cancer, and discuss why we believe many of these studies are predisposed to show minor or no effects, and then suggest approaches that we hypothesize will provide more conclusive evidence on whether stress significantly affects long-term cancer outcomes in humans.

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## Sympathetic nervous system

(SNS). Part of the autonomic nervous system that is involuntarily activated by stressors (for example, a dangerous or stressful situation) and orchestrates the 'fight or flight' response through adrenergic innervation of the adrenal medulla and of various organs (for example, the heart) through systemic and local release of adrenaline and noradrenaline, respectively.

## Hypothalamic–pituitary–adrenal (HPA) axis

A neuroendocrine system with negative feedback that increases systemic glucocorticoid (for example, cortisol) levels in various circumstances, including stressful conditions. Hypothalamic corticotropin-releasing hormone (CRH) elevates systemic release of adrenocorticotrophic hormone (ACTH) from the anterior pituitary, which triggers the release of glucocorticoids from the adrenal cortex, which also trigger negative feedback through the pituitary and hypothalamic levels.

## Damage-associated molecular patterns

Endogenous host-derived molecules that are released by damaged and dying cells. They are recognized by pattern recognition receptors on numerous cells, which lead to migration and activation of various immune cells and consequent innate and adaptive immune responses.

## Stress and stress responses

Hans Selye in 1956 (REF.<sup>14</sup>) described stress as a response of the body to the demands made upon it in an attempt to return to homeostasis. Meeting the demands of life, spanning from day to day tasks to major threats such as the diagnosis and treatment of cancer, requires mobilization of metabolic energy to sustain necessary physiological adaptive responses. This is achieved by activation of the sympathetic nervous system (SNS) and the hypothalamic–pituitary–adrenal (HPA) axis, leading to local and systemic secretion of adrenergic factors from sympathetic nerve endings and the adrenal medulla (mostly noradrenaline (also known as norepinephrine) and adrenaline (also known as epinephrine), respectively), the release of glucocorticoids (such as cortisol) from the adrenal cortex, and the secretion of opioids, oxytocin and other stress mediators (FIG. 1).

The stress responses described above are initiated by the central nervous system (CNS) following processing of various stimuli, including physiological inner-body responses to various conditions, such as tissue damage (including during surgery and under anaesthesia), or being subjected to low temperature; external stimuli, such as being attacked by an assailant with a weapon or being informed of having cancer; or ongoing CNS activities, resulting from being anxious or ruminating about financial insecurity, social isolation, interpersonal relationships or having cancer (FIG. 1). Notably, both depression and social isolation involve activation and/or dysregulation of the HPA axis, and are characterized by a pro-inflammatory state<sup>15,16</sup>, which triggers similar pathways to stress responses (discussed below). Last, stress and depression promote each other<sup>17</sup>, and most animal models of depression are based on stress exposure<sup>18</sup>.

Stress can be both beneficial and deleterious (BOX 1). The effects of stress on the capacity of an organism to cope with challenges typically follow an inverted U shape<sup>19</sup> — when the intensity, duration or nature of the stressor is moderate, stress facilitates adaptive natural changes, but when stress exceeds the resources of the individual to cope, and becomes 'toxic stress', the risk for disease increases<sup>20</sup>. McEwen and Stellar defined allostasis as the naturally occurring continuous adaptations towards different homeostatic states<sup>21</sup>. When allostasis becomes strenuous, and the allostatic load increases to the point of overload, patients are at greater risk<sup>21</sup>.

Notably, the intensity and duration of stress responses to internal or external stimuli markedly differ between individuals, and depend on physiological factors, including genetic and developmental variations<sup>19</sup>, and physical fitness (BOX 2); individual psychosocial characteristics, including perceived social support<sup>22</sup>, perceived ability to cope<sup>23</sup> and other personal traits; and the characteristics of the stressful life events previously experienced<sup>24–26</sup>, including childhood adversity<sup>27</sup>. It follows that stressors such as cancer diagnosis, treatment and survivorship are likely to be differentially experienced by patients, provoking different stress responses. Thus, stress-management therapies, behavioural or pharmacological, should be individually tailored. Additionally, understanding specific physiological mechanisms mediating deleterious (or beneficial) effects of stress responses may

point to effective downstream pharmacological therapeutic approaches, which may also surpass individual differences at higher psychological/cognitive levels.

## Critical periods in cancer progression

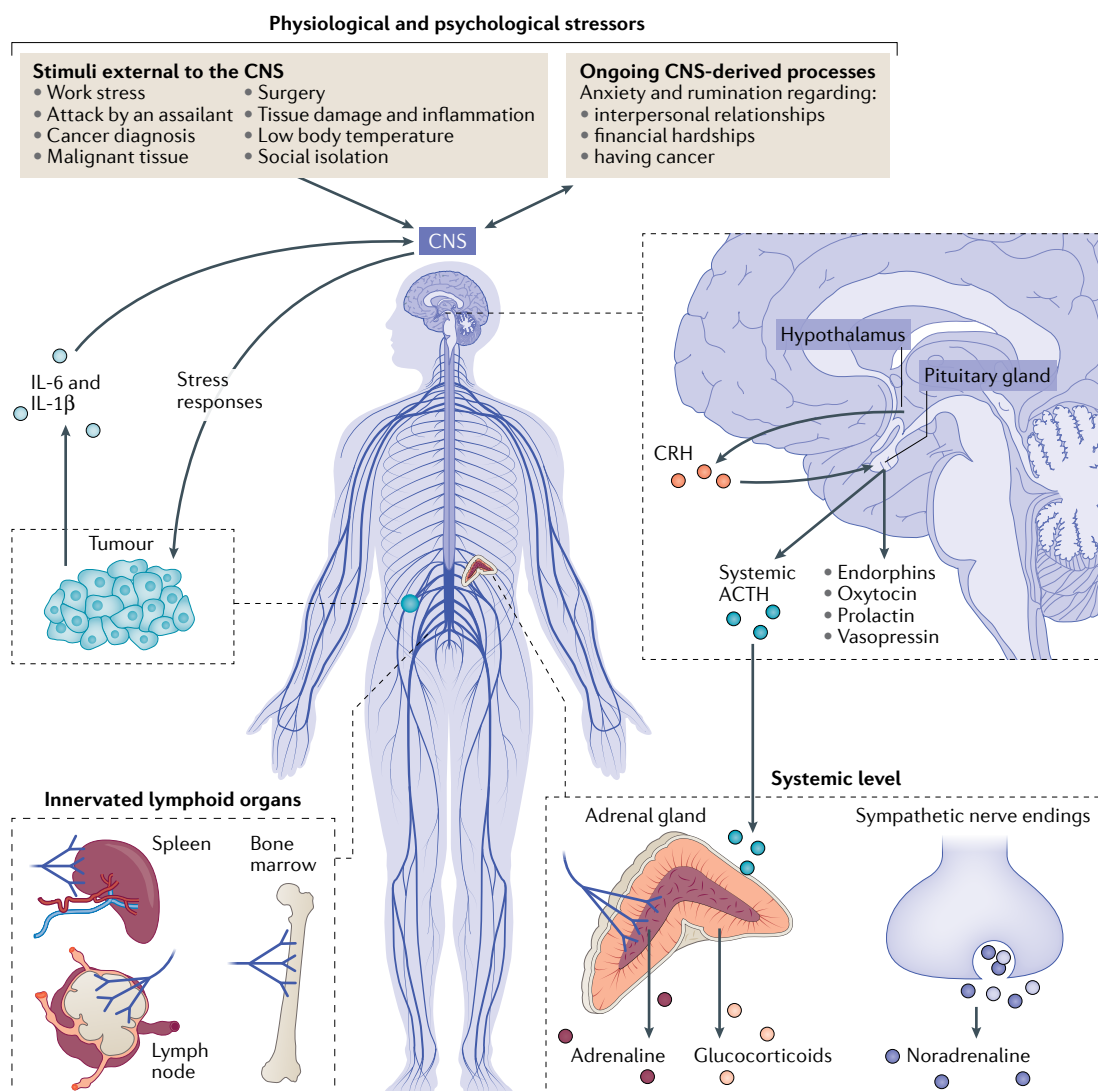
Normal cells transform into malignant cells through acquisition of unique characteristics with evolutionary advantages, known as the 'hallmarks' of cancer<sup>28,29</sup>. These characteristics include resistance to apoptotic signals, independence from external growth signals, the capacity to attract vascularization, evasion of immune destruction and the acquisition of invasive properties into distant organs with a permissive microenvironment to form metastases. Importantly, along this transformation, pre-malignant or malignant foci may be eliminated, may become dormant or slowly progressing<sup>30</sup>, or may advance to a clinical manifestation.

Theoretically, some phases may be more critical along this multimodal non-linear process. Examples include activation of the 'angiogenic switch' that enables increased growth or escape from dormancy<sup>31</sup>; initial interactions with immune cells following neo-vascularization and/or release of damage-associated molecular patterns<sup>32</sup>; the passage of circulating tumour cells through pulmonary or hepatic capillaries, where highly active marginating natural killer cells recognize and eliminate such aberrant cells<sup>33–35</sup>; survival of circulating tumour cells in the circulation and extravasation into new organs<sup>36</sup>; and the capacity of a micrometastasis to grow independently of the primary tumour<sup>37</sup>.

Stress may have greater impact during such potential critical phases. Moreover, whether stress will exacerbate or mitigate malignant processes may depend on the phase of malignant progression, specific tumour characteristics and the spectrum of stress responses. Also, immune system–tumour interactions may either impair or promote tumour growth<sup>38</sup>, and stress hormones can regulate both processes<sup>7,9</sup>. Thus, interactions between stress and cancer are expected to be non-linear, and the impact of stress could depend on the phase of cancer progression.

Hypothetically, an acute or chronic stress episode that is synchronized with a critical phase may bear a greater impact on cancer progression than non-synchronized episodes. Studies in animal models, more than clinical or epidemiological studies, can focus on a critical phase, employing specific tumour types, and/or stress paradigms, and thus maximize our ability to observe the potential impact of stress. For example, stressing animals shortly before and after intravenous tumour cell inoculation maximizes the deleterious impact of stress on the capacity of marginating pulmonary natural killer cells to prevent experimental lung metastasis<sup>33,39,40</sup>. In breast cancer mouse models, chronic stressors did not affect growth of primary tumours but did promote their dissemination and metastatic growth<sup>41,42</sup>. Last, subjecting mice to chronic social isolation before mammary tumour inoculation exerted no effects on primary tumour growth, whereas if initiated when tumours were palpable, primary tumour growth was increased<sup>43</sup>.

In the clinical setting, some critical phases cannot be recognized but others, especially those related to cancer



**Fig. 1 | Stress responses and reciprocal stress–cancer interactions.** Physiological and psychological stressors including stimuli external to the central nervous system (CNS), such as being informed of having cancer, undergoing surgery or the presence of malignant tissue and its related inflammation, and ongoing CNS-derived processes (for example, anxiety and rumination about cancer) are perceived and processed by the CNS and trigger stress responses. Consequently, the pituitary gland releases endorphins, oxytocin, prolactin, vasopressin, adrenocorticotropic hormone (ACTH) and other stress mediators, and activation of the hypothalamic–pituitary–adrenal (HPA) axis through hypothalamic corticotropin-releasing hormone (CRH) and systemic ACTH release leads to secretion of glucocorticoids (for example, cortisol) from the adrenal cortex. Simultaneously, the CNS activates the sympathetic nervous system (SNS), leading to secretion of adrenergic factors from the adrenal medulla (mostly adrenaline) and sympathetic nerve endings (mainly noradrenaline). The latter also innervate lymphoid organs (for example, spleen and lymph nodes), bone marrow and various organs. These stress factors promote most hallmarks of cancer through impacting the malignant tissue, its microenvironment, immunity, lymphatic flow and distant potential pre-metastatic niches (FIG. 2). Malignant tissue can facilitate stress responses through local and systemic inflammation (for example, through interleukin-6 (IL-6) and IL-1 $\beta$ ) that affects the CNS, dysregulates HPA axis activity<sup>220,221</sup> and promotes depression, sleep disturbances and cancer-related fatigue. Overall, CNS-initiated stress responses may lead to exacerbated tumour growth and spread, and to peripheral stress–inflammatory–cytokine responses, which feed back to the CNS, altering cognition and mood, and facilitating stress responses, creating a vicious cycle.

treatment, are known to impact cancer progression (BOX 3), and can perhaps be exploited to mitigate the effects of stress on cancer progression.

### Mechanisms of stress impacts on cancer

As briefly reviewed below, a vast body of literature indicates that stress can promote most hallmarks of cancer<sup>2</sup>, and mechanisms mediating these effects by specific

stress hormones, their receptor systems and intracellular molecular mechanisms have been identified (reviewed in REFS<sup>3,4,6,44</sup>). We herein discuss and refer to tumour initiation as transformation from non-malignant to malignant tissue, in contrast to tumour progression that follows this transformation, although most hallmarks of cancer can affect both initiation and progression of the disease. We present causative findings from

## Box 1 | Acute and chronic stress

Acute stress is defined as lasting minutes to hours, whereas chronic stress can last days, weeks, months or longer<sup>271</sup>. A short-term transient stress response can be adaptive, as part of the 'fight or flight' response, where sympathetic nervous system (SNS) and hypothalamic–pituitary–adrenal (HPA) axis activation increases the heart rate, blood pressure and glucose availability. Such stress responses can also promote the release of pro-inflammatory cytokines (for example, interleukin-6 (IL-6) and IL-1 $\beta$ ) and trafficking of leukocytes to the skin following stress cessation<sup>272,273</sup>, potentially to allow skin pathogen resistance in the case of injury<sup>271</sup>. By contrast, long-lasting or repeated stress exposures can lead to HPA axis dysregulation, glucocorticoid resistance and/or insensitivity to HPA axis negative feedback<sup>274</sup>. These may lead to chronic inflammation secondary to disrupted HPA axis-induced inhibition of pro-inflammatory responses<sup>274</sup>. Nevertheless, chronic elevated levels of glucocorticoids contribute to immunosuppression<sup>275</sup>. Moreover, animal studies have demonstrated that both acute and chronic stress paradigms can suppress immunity<sup>40,273</sup> and promote certain anti-inflammatory responses, such as decreased plasma IL-12 levels<sup>276</sup>.

Notably, the distinction between acute and chronic stress is often ambiguous. Chronic stress paradigms in animals are often based on repeated<sup>41</sup> or alternating<sup>85</sup> acute stressors, rather than continuous stressors. Furthermore, there is no unified definition of acute or chronic stress<sup>82,85,92,135</sup>, with 3–5 consecutive days of repeated acute stressors defined both as acute<sup>92</sup> and as chronic<sup>85</sup> stressors. Also, continuous chronic social isolation was found to increase reactivity to acute restraint stress<sup>67,137</sup>, demonstrating mutual interdependence between acute and chronic stress. In humans, acute events can generate a chronic threat perception and/or chronic stress responses<sup>277</sup>, especially given pre-event anticipation and post-event ruminations<sup>27</sup>. In the context of cancer treatment, the overlapping nature of acute and chronic medical and psychological stressors, and the psychological consequences of these events, may mask the distinction between acute and chronic stress and their impact on cancer progression. Moreover, some naturally adaptive responses to acute stress, such as redistribution of leukocytes to the skin at the expense of internal organs, may increase the risk for internal organ metastasis, as indicated by animal studies employing acute stressors<sup>40,133</sup>. Thus, the intricacies of acute and chronic stress responses in the context of cancer progression and treatment suggest caution in making any generalizations.

### Catecholamines

A family of molecules that are characterized by a catechol and an amine group in their chemical structure, and function as neurotransmitters and hormones within the body. These include dopamine, noradrenaline and adrenaline, all of which are synthesized from the amino acid tyrosine.

### Restraint stress

An experimental stress paradigm, where the animal is placed in a confined space (a tube-shaped apparatus perforated for air exchange) that prevents free movement but does not press or induce pain to the animal. Such restraint can last minutes to hours and can be repeated daily for several weeks as a chronic stress paradigm.

### Sympathetic denervation

Refers to experimental methods for ablation of sympathetic nerves (also called sympathectomy), by either surgical cut of sympathetic nerve fibres or chemical ablation (for example, using 6-hydroxydopamine).

animal studies, which often are followed by parallel clinical findings, complementing each other in terms of methodological robustness and clinical relevance.

### Cancer initiation

**DNA damage.** Specific stress factors have been shown to cause DNA damage and jeopardize DNA repair, potentially facilitating malignant transformation. Specifically, in a mouse fibroblast cell line, serum derived from stressed mice, or adrenaline, noradrenaline and cortisol (each factor alone as well as synergistically when combined), increased DNA damage and/or reduced DNA repair following UV irradiation<sup>45</sup>. In murine and human non-cancer cell lines,  $\beta$ -adrenergic receptor ( $\beta$ -AR)-mediated generation of reactive oxygen species and  $\beta$ -arrestin–MDM2-dependent p53 degradation increased DNA damage and inhibited DNA repair<sup>46</sup>. Corresponding in vivo studies confirmed that chronic stress induces these two  $\beta$ -AR-mediated processes<sup>47</sup>, and that glucocorticoid-mediated response can also cause MDM2-dependent p53 downregulation and increase resistance to apoptosis following ionization irradiation<sup>48</sup>. In humans, several studies indicated that psychological stress is associated or causatively linked to increased DNA damage<sup>49</sup>, and several human cancer cell lines exhibited accelerated DNA damage in vitro following  $\beta$ -adrenergic and glucocorticoid signalling<sup>50–52</sup>, in part through activation of the ATR–p21 pathway<sup>52</sup>. Nevertheless, it should be noted that DNA damage alone

is insufficient to cause tumour initiation, as mutations need to be maintained and accumulated across repeated cell divisions, and should lead to acquisition of resistance to apoptosis and to increased proliferation, among other characteristics.

**Oncogenic viruses.** Thirteen to 15% of human cancer incidence is attributed to carcinogenic infections<sup>53,54</sup>, and stress can also increase the risk for cancer initiation by promoting the prevalence and outbreak of oncogenic viruses. Following in vitro infection of various human cell lines, major oncogenic human viruses were shown to be reactivated by either glucocorticoids or catecholamines, including human papillomaviruses (HPVs), Epstein–Barr virus, Kaposi sarcoma-associated herpesvirus and hepatitis B and C viruses<sup>55</sup>. Additionally, stress hormones were shown to stimulate oncogene expression in human cells infected with oncogenic viruses, as well as to suppress expression of type I interferons (IFN $\alpha$  and IFN $\beta$ ) in leukocytes, impairing antiviral immunity<sup>55–57</sup>. In humans, academic examination stress in cadets, and/or activation of the SNS or HPA axis, was associated with reactivation of latent oncogenic viruses<sup>58,59</sup>; higher levels of perceived stress were associated with impaired HPV-specific T cell responses in women with cervical dysplasia<sup>60</sup>; and loss of a child predicted increased risk for HPV-associated cancers in a cohort of more than four million parents in Sweden<sup>61</sup>.

**Tumorigenesis.** Several in vivo animal studies assessed the effects of stress on actual tumorigenesis, rather than on interim indices, such as DNA damage or reactivation of oncogenic viruses. Repeated restraint stress<sup>48,62</sup>, social isolation<sup>63</sup> and cold ambient temperature<sup>64</sup> promoted carcinogen-induced tumorigenesis. In transgenic models of spontaneous cancer, repeated restraint stress increased pancreatic tumorigenesis through  $\beta_2$ -AR signalling<sup>65</sup>, whereas sympathetic denervation decreased tumorigenesis in a prostate cancer model<sup>66</sup>. However, in such models that are based on accelerated induction of cancer, it is hard to distinguish between effects of stress on tumour initiation and its effect on tumour progression, as the time course of stress largely overlaps with both initiation and progression periods<sup>48,65,67,68</sup>. Thus, stress can potentially exacerbate the effects of carcinogenic exposure, yet it is unclear whether stress is a significant factor in tumour initiation in the absence of known exposure to carcinogens.

### Cancer progression

**Direct effects on tumour cells.** Stress hormones, secreted systemically or released locally in the tumour micro-environment from sympathetic nerve endings, immune cells<sup>69,70</sup> or tumour cells<sup>71–73</sup>, can directly affect tumour cells, promoting their malignant characteristics. Specifically, noradrenaline and adrenaline were shown in vitro to promote tumour cell proliferation<sup>74–76</sup>, survival (anti-apoptosis)<sup>74,75,77</sup>, migration<sup>74,78,79</sup>, invasion<sup>74,78–81</sup>, epithelial–mesenchymal transition (EMT)<sup>42,78,82,83</sup> and production of prostaglandins<sup>76,79</sup> and matrix metalloproteinases (MMPs)<sup>76,80,81</sup> (FIG. 2). Accordingly, behavioural or physiological stressors (for example, social



### Prostaglandin receptors

A class of cell surface G-protein-coupled receptors that bind different prostaglandins and are expressed on various cell types, including immune cells; for example, prostaglandin  $E_2$  binds to the prostaglandin  $E_2$  receptor 1–4 subtypes.

### T helper 1 cell

( $T_H1$  cell). A CD4<sup>+</sup> T cell that participates in the pro-inflammatory type 1 or cellular immune response against intracellular pathogens and malignant cells. Naive T cells are differentiated into the type 1 phenotype following exposure to interleukin-12 (IL-12), and are known for the secretion of interferon- $\gamma$  (IFN $\gamma$ ), which is also involved in the effector functions of cytotoxic T cells.

confrontation, restraint and surgery) in animal models were shown to increase tumour growth and metastasis through activation of tumour  $\beta$ -AR, as indicated by their specific pharmacological<sup>41,74,80,84–86</sup> or molecular<sup>87,88</sup> blockade, or by genetic knockout<sup>84</sup>.

Recent studies have indicated the contribution of tumour innervation to tumour progression<sup>89</sup>. Tumours can secrete neuronal growth factors, increasing sympathetic tumour innervation. This creates a feedforward loop that promotes cancer progression under stress-induced sympathetic activation, as a result of higher tumoural noradrenaline levels<sup>65</sup>. Correspondingly, numerous human cancers were found to express  $\beta$ -AR<sup>65,74,75,78,79,81,83,90</sup>, and their higher expression<sup>74,75,78,79,83</sup> or higher tumour noradrenaline<sup>91</sup> and/or plasma adrenaline<sup>82</sup> levels were correlated with larger tumour size, advanced stage, lymph node metastasis and/or reduced survival in several cancer types. Interestingly, low social support in patients with ovarian cancer predicted higher tumour levels of noradrenaline<sup>91</sup>.

Behavioural stress can also promote tumour growth through glucocorticoid secretion<sup>48,92</sup>, and synthetic glucocorticoid receptor (GR) agonists (for example, dexamethasone) promoted metastasis and reduced survival in xenograft and syngeneic breast cancer models<sup>93</sup>. In patients with breast cancer, higher tumour expression levels of GR and GR-regulated kinases predicted poorer survival<sup>93,94</sup>.

## Box 2 | Physical exercise, stress and cancer

Physical exercise exerts a challenge to whole-body homeostasis, promoting extensive adaptations of cells, tissues and organs<sup>278</sup>. Moderate physical exercise is known to improve cardiometabolic indices, to increase cognitive performance and to improve numerous health conditions and support their treatment, including cancer<sup>279</sup>. Physical exercise increases the levels of stress hormones (for example, adrenaline, endorphins and cortisol) for the duration of the exercise, blunts hormone responses to stress<sup>280,281</sup> and modulates inflammatory status and cytokine levels during exercise (for example, increased interleukin-6 (IL-6), IL-10 and IL1R $\alpha$ , but not TNF and IL-1 $\beta$ )<sup>282</sup>. In the context of cancer, physical exercise was shown to have beneficial impact on quality of life, fatigue, anxiety, depressive symptomatology and psychological distress<sup>283–287</sup>. The effect of exercise on inflammation is complex. In the general population, physical exercise is generally associated with reduced inflammation<sup>282</sup>, whereas in patients with cancer this association is more limited<sup>288</sup>. Importantly, prospective correlational studies indicated that physically active patients have significantly lower mortality rates than non-active patients<sup>289,290</sup>. Interestingly, whereas stress responses exert numerous pro-tumorigenic effects (as reviewed herein), physical exercise-induced stress factors exhibit antitumorigenic properties<sup>291</sup>. For example, in preclinical studies, exercise-conditioned serum, derived from healthy humans and patients with cancer, had growth-inhibitory effects on breast cancer cell lines in vitro and in vivo<sup>292</sup>. Moreover, mice subjected to voluntary physical exercise had attenuated tumour growth and enhanced antitumour activity via  $\beta$ -adrenergic signalling<sup>292–294</sup>. Hypothesized explanations for the apparent contradictory beneficial and deleterious effects of  $\beta$ -adrenergic signalling include the rapid and transient increase and decrease of adrenergic responses to exercise; inhibited stress responses following physical exercise; and the rapid exercise-related mobilization of cytotoxic immunocytes (for example, CD8<sup>+</sup> T cells, natural killer cells)<sup>295</sup> to the circulation, as opposed to stressors and their aftermath that induce immunosuppression. Additionally, physical exercise was shown to exert the production of dihydroxyphenylalanine (DOPA) and dopamine (as part of the catecholamine response)<sup>296</sup> that were reported to antagonize tumour progression<sup>10</sup>, whereas these responses are generally not induced by stressors. Overall, these results warrant further studying of the mechanisms by which physical exercise improves psychological indices, physical adaptation to stress and malignant conditions, and devising suitable exercise regimens for patients with cancer to potentially improve short-term and long-term outcomes.

**Angiogenesis and lymphangiogenesis.** In vitro findings indicated that noradrenaline and adrenaline increase tumour cells' expression and secretion of several angiogenic factors, including vascular endothelial growth factor (VEGF), interleukin-6 (IL-6) and IL-8 (REFS<sup>81,95–97</sup>), and that noradrenaline-mediated angiogenesis is reinforced following direct contact between tumour cells and endothelial cells<sup>98</sup>. In stressed nude mice orthotopically implanted with human ovarian carcinoma cells,  $\beta_2$ -AR–cyclic AMP (cAMP)–protein kinase A (PKA) signalling increased tumour expression of VEGF, and tumour vascularization and growth<sup>87</sup>. Similar findings were confirmed in pancreatic cancer<sup>99</sup>, colorectal cancer (CRC)<sup>100</sup> and breast cancer<sup>41,101</sup> models. Stress-induced  $\beta$ -AR signalling also inhibited the anti-angiogenic factor thrombospondin 1 (TSP1) in prostate cancer xenografts through epigenetic modulation<sup>102</sup>. In patients with ovarian carcinoma, lower social well-being and elevated distress or depressive symptoms correlated with higher plasma and tumour VEGF levels<sup>103,104</sup>, and higher ascites and plasma IL-6 levels<sup>105,106</sup>.

Tumour lymphatic vessel density and lymphangiogenic growth factors are associated with metastases and with reduced survival in patients with cancer<sup>107</sup>. Chronic restraint stress in mice, through  $\beta$ -AR signalling, increased expression of the lymphangiogenic factor VEGFC in tumour and stromal cells, and increased expression of cyclooxygenase 2 (COX2; also known as PTGS2) in tumour-associated macrophages (TAMs). These changes led to elevated lymphatic vessel density and increased metastasis<sup>108</sup>. In patients with cancer, acute blockade of SNS activity reduced lymph flow in patients with cervical carcinoma<sup>108</sup>, and breast tumours in socially isolated women exhibited increased density of lymphatic vessels<sup>109</sup>.

**Immunomodulation and inflammation.** Stress has been shown to promote both inflammation and immune evasion<sup>8</sup>. Most immune cells express  $\beta$ -ARs<sup>110</sup>, prostaglandin receptors<sup>111</sup> and GRs<sup>44</sup>, and the effects of stress on their activity and distribution have been extensively studied in animal models and in patients with cancer<sup>7–9,110</sup>.

In murine models, natural killer cell activity against tumour cells was suppressed by stress-induced  $\beta$ -adrenergic signalling or  $\beta$ -adrenergic agonists<sup>33,40,112,113</sup>, and a stress-induced increase in lung metastases was shown to be mediated by suppression of natural killer cells<sup>114</sup>. In patients with ovarian cancer, lower social support and higher distress correlated with lower natural killer cytotoxicity<sup>115</sup>. Stress was also shown to induce a shift from T helper 1 cell ( $T_H1$  cell)-type to T helper 2 cell ( $T_H2$  cell)-type cytokine production, to increase tumour growth in mouse models of CRC<sup>116</sup> and squamous cell carcinoma<sup>62</sup>, as well as to increase tumour growth through  $\beta$ -AR-mediated suppression of CD8<sup>+</sup> T cells in mammary and melanoma mouse models<sup>84</sup>. Correspondingly, in patients with ovarian carcinoma, depressed and anxious mood correlated with a reduced  $T_H1$  cell/ $T_H2$  cell-type cytokine ratio<sup>117</sup>. Additionally, in mouse models, a stress-induced  $\beta$ -adrenergic response promoted tumour growth by upregulation of suppressive

### Box 3 | Critical time frames during cancer treatment

Along the course of cancer treatment, there are recognized critical phases where susceptibility to the impacts of stress may be heightened. These include the surgical removal of the primary tumour, and neoadjuvant and adjuvant therapies. Specifically, during the short perioperative period (days before and after surgery), surgical excision of the malignant mass may increase shedding of tumour cells to the circulation<sup>297,298</sup>, terminate primary tumour-related secretion of anti-angiogenic factors<sup>299,300</sup> and induce the release of growth factors<sup>301,302</sup>. These processes cumulatively or synergistically increase the risk of metastatic disease<sup>198,199</sup>. Moreover, stress and inflammatory responses are elevated as a result of psychological distress, tissue damage, hypothermia, blood transfusions, pain and specific analgesic/anaesthetic approaches<sup>198,199</sup>. These neuro-endocrine responses, especially catecholamine and prostaglandin signalling, suppress antitumour immunity<sup>9,303</sup>, and directly facilitate progression of residual disease, as elaborated in the main text. Most importantly, as the short perioperative period holds a delicate balance between pro-metastatic and anti-metastatic processes, stress responses during this time can tilt the balance towards the pro-metastatic direction, creating a 'snowball effect' that impacts long-term cancer outcomes<sup>186</sup>. Indeed, several clinical perioperative events (for example, anastomosis leak and/or secondary surgery) or specific medical routines (for example, use of the sedative dexmedetomidine) were associated with worse long-term cancer outcomes<sup>304</sup>, and animal studies provided causative evidence that such events can increase the deleterious impacts of stress on cancer metastasis<sup>305</sup>. Additionally, a recent study in rodents reported that the effects of pre-surgical behavioural stress exacerbate the deleterious effects of surgery on lung metastasis<sup>133</sup>.

The peri-adjuvant time frame also constitutes a critical period of cancer progression. Adjuvant therapies and their side effects are accompanied by psychological distress<sup>306</sup>, induce inflammatory responses<sup>307</sup> and can promote tumorigenic and metastatic processes<sup>308</sup>. For example, the chemotherapies cisplatin and paclitaxel activate the pro-inflammatory nuclear factor- $\kappa$ B (NF- $\kappa$ B) pathway, inducing the expression of various pro-tumorigenic and pro-metastatic factors such as interleukin-6 (IL-6) and IL-8, and promoting angiogenesis and tumour cell proliferation, survival and epithelial-mesenchymal transition (EMT)<sup>307</sup>. Adjuvant therapies can lead to selection of drug-resistant tumour clones, and to host-derived responses that promote cancer recurrence<sup>308</sup>. Thus, as adjuvant therapies have both pro-tumour and antitumour effects, and as stress during cancer therapy can impair its efficacy (as discussed in the main text), stress may have greater impact during the peri-adjuvant time frame.

As the short perioperative and the peri-adjuvant time frames are characterized by excessive stress and inflammatory responses and by accelerated tumour progression, they could be exploited therapeutically for anti-metastatic approaches, and specifically interventions that reduce stress and inflammation.

#### T helper 2 cell

(T<sub>H</sub>2 cell). A CD4<sup>+</sup> T cell that participates in type 2 or humoral immune response against extracellular pathogens (for example, helminths) and allergens. Naive T cells are differentiated into a type 2 phenotype following exposure to interleukin-4 (IL-4), and are known for the secretion of IL-4, IL-13 and IL-5, and promotion of the production of antibodies.

#### $\beta$ -Blockers

A class of drugs with antagonistic activity towards  $\beta$ -adrenergic receptors ( $\beta$ -ARs). The drugs vary in specificity to the different  $\beta$ -ARs ( $\beta_1$ -AR,  $\beta_2$ -AR and  $\beta_3$ -AR) and are classified as selective or non-selective to a certain receptor subtype.

immune cells such as myeloid-derived suppressor cells (MDSCs) and regulatory T cells<sup>62,84,100,118</sup>, whereas in patients with breast cancer, higher levels of stress correlated with reduced numbers of circulating MDSCs<sup>119</sup>.

With respect to inflammation, stress-induced  $\beta$ -adrenergic signalling in preclinical studies was shown to promote COX2 expression and prostaglandin secretion in both tumour cells and TAMs<sup>41,79,108</sup>, to stimulate secretion of pro-inflammatory cytokines (for example, IL-6)<sup>95,97</sup> and to increase tumour recruitment of macrophages and their M2 polarization<sup>41,90,120,121</sup>. Correspondingly, in patients with cancer, social isolation correlated with upregulation of M2 polarization in breast tumours<sup>109</sup>, higher levels of depression were associated with higher levels of prostaglandins in ovarian tumours<sup>79</sup>, and tumour expression levels of genes encoding  $\beta_2$ -AR and prostaglandins predicted reduced survival<sup>79</sup>.

**Metastasis.** Metastases are promoted by many of the aforementioned mechanisms, as well as by additional stress-induced processes. For example, in mice, stress-induced  $\beta$ -AR activation promoted migration of circulating tumour cells to the bones, through increased

expression of receptor activator of nuclear factor- $\kappa$ B ligand (RANKL) by bone marrow stem cells (BMSCs)<sup>122</sup>, or to the lungs by CC-chemokine ligand 2 (CCL2)-CC-chemokine receptor 2 (CCR2)-mediated attraction of macrophages<sup>85</sup>, consequently forming pre-metastatic niches and increasing organ-specific metastasis. Additionally, stress increased tumour cell EMT<sup>42,82,83</sup>, tumour and stromal cell secretion of MMPs<sup>41,74,80,99</sup> and tumour cell resistance to anoikis<sup>77</sup>, promoting malignant cell detachment, invasiveness and survival in the circulation<sup>123</sup>. In patients with breast and ovarian cancer, perceived stress, depressive symptoms or social isolation predicted higher tumour expression of EMT-related genes<sup>109,124</sup>, and higher MMP9 levels in tumour cells and/or TAMs<sup>104</sup>. Importantly,  $\beta$ -AR blockade reduced stress-induced metastasis in many murine models, of both experimental and spontaneous metastases<sup>33,41,74,85,108,122,125</sup>. Correspondingly, in patients with gastric and lung cancer, tumour  $\beta$ -AR expression levels correlated with lymph node metastasis<sup>74,126</sup>, and incidental use of  $\beta$ -blockers was associated with decreased metastasis or recurrence in patients with breast and ovarian cancer<sup>108,127,128</sup> and with improved survival in melanoma and breast cancer<sup>129,130</sup>, but not in lung and ovarian cancer<sup>131,132</sup>. These diverse outcomes are expected given differences between the indices studied (for example, metastasis versus survival), diverse cancer types and the uncontrolled settings of correlational studies, and call for randomized controlled trials (RCTs) to test the effects of  $\beta$ -blockers on long-term cancer outcomes.

**Acute and chronic stressors.** Although most animal studies report that stress, whether acute or chronic, promotes primary tumour growth and metastasis, a few studies report that stress can decrease primary tumour growth. For example, several paradigms of acute stress were reported to increase primary tumour growth and metastasis in rodents, including restraint stress<sup>92</sup>, 16-h tilt-light stress<sup>133</sup>, 30–60 min of intermittent swim stress<sup>39,40,113</sup>, laparotomy<sup>100,114,133</sup> or 7 h of social confrontation stress<sup>134</sup>, whereas other studies showed that acute restraint stress<sup>135</sup> and foot shock stress<sup>136</sup> can inhibit primary tumour growth. Heterogeneity of the acute stressors, tumour models, animal species and phase of tumour progression during stress exposure may underlie this apparent inconsistency (as discussed above). With respect to chronic stress, and examining a more standardized setting of chronic social isolation in breast cancer models, stress exposure increased<sup>67,137</sup>, decreased<sup>168,138</sup> or had transient<sup>43</sup> effects on primary tumour growth. Although there are physiological differences between acute and chronic stress (BOX 1), comparison between acute and chronic restraint stress showed that whereas the stress duration had differential effects on spleen T lymphocytes, neither acute nor chronic stress affected the growth of primary mammary tumours but both increased blood vessel density in metastatic foci<sup>101</sup>. Additionally, chronic social isolation, but not chronic restraint, reduced survival of mammary tumour-bearing mice<sup>101</sup>. As elaborated in BOX 1, there is ambiguity regarding the definitions of acute and chronic stressors, and some adaptive characteristics of

### Tilt–light stress

An experimental stress paradigm in which the home cage of rodents is placed in a lit room in a 45° tilted position, starting before the onset of the animals' dark period, resulting in reduced available floor space and disruption of the dark–light cycle.

### Swim stress

An experimental stress paradigm where a weight is attached to the tail of rodents (usually rats, up to 2.5% of total body weight), which are then placed in a room temperature water tank for few minutes, followed by a rest period. This swim–rest cycle is usually repeated several times.

acute stress responses in the natural setting may promote cancer progression. Importantly, no generalization can be drawn regarding stress chronicity and cancer progression, and other aspects of stress–cancer interactions may be more critical. Overall, the majority of animal studies report that stress promotes primary tumour progression, rather than inhibits it. The impact of stress on metastasis seems even more consistent, with the great majority of studies reporting increased pro-metastatic processes, and few reporting no impact.

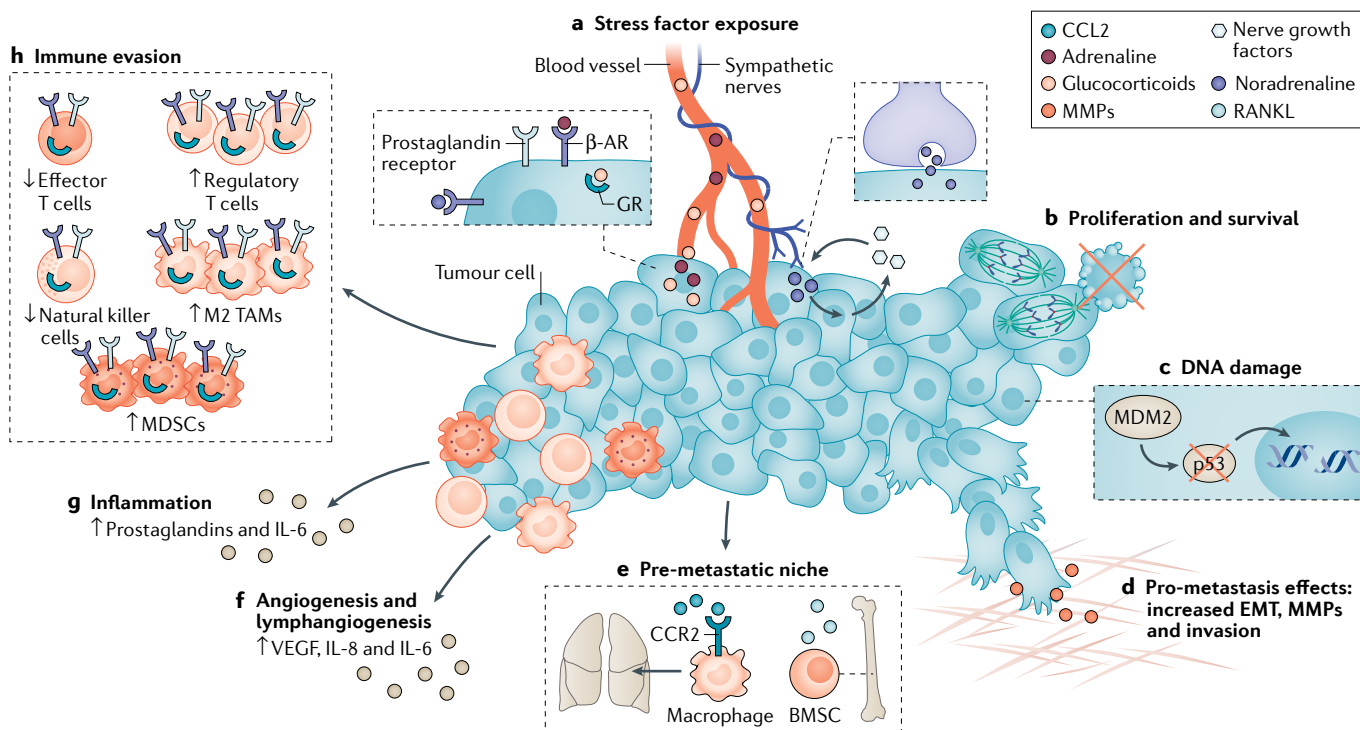
In summary, the effects of stress, acute or chronic, on tumour progression and metastasis are robust; are mediated by  $\beta$ -adrenergic signalling; are mediated to a lesser degree by HPA axis signalling<sup>114</sup>; and occur through affecting tumour cells, and their microenvironment, including immune and stromal cells (FIG. 2; TABLE 1). Notably,  $\beta$ -AR signalling that promotes tumour progression corresponds with natural adrenergic effects on healthy/non-malignant tissue, including adrenergic effects on EMT<sup>139,140</sup>, inflammation<sup>141–143</sup> and

angiogenesis<sup>144,145</sup> (not in the context of cancer). Last, whereas rodent models and in vitro cell culture provide causal evidence for specific links between stress responses and tumour progression, findings from studies in patients are mostly correlative, with the exception of a few intervention studies reviewed below.

## Epidemiological studies

### Stress and cancer incidence

A comprehensive meta-analysis of 142 prospective studies published in 2008 (REF.<sup>146</sup>) (average sample size of 87,000 people per study) indicated that psychosocial stress predicts a 6% increase in cancer incidence (hazard ratio = 1.06; 95% CI 1.02–1.11,  $P=0.005$ ). Of note, depression was a major factor in this effect, rather than stressful life events. However, the meta-analysis identified a significant publication bias, suffered from marked heterogeneity in the outcomes of the included studies and was criticized for meta-analytic methodological flaws<sup>147</sup>. Moreover, 76% of the studies reported a



**Fig. 2 | Effects of stress on the tumour and its microenvironment.**

Malignant tissue is exposed to systemic stress factors, including adrenaline, noradrenaline and glucocorticoids (for example, cortisol in humans), and to locally released noradrenaline through sympathetic tumour innervation (part a). Tumours can also release nerve growth factors that increase their sympathetic innervation and noradrenaline exposure, creating a feedforward loop. Through membrane-bound  $\beta$ -adrenergic receptors ( $\beta$ -ARs), which bind adrenaline and noradrenaline, and intracellular glucocorticoid receptors (GRs), all of which are expressed by tumour, immune and stromal cells, stress factors promote most hallmarks of cancer. Tumour cell proliferation and resistance to cell death are increased (part b). In addition, activation of  $\beta$ -ARs and GRs also induces activation of the E3-ubiquitin ligase MDM2 and consequent degradation of p53, which leads to impaired genome maintenance and accumulation of DNA damage (part c). Stress factors promote invasion and metastasis by inducing tumour epithelial–mesenchymal transition (EMT) and the release of matrix metalloproteinases (MMPs) (part d). Furthermore, activation of  $\beta$ -ARs promotes the formation of

organ-specific pre-metastatic niches through processes such as CC-chemokine ligand 2 (CCL2)–CC-chemokine receptor 2 (CCR2)-mediated attraction of macrophages to the lung and receptor activator of nuclear factor- $\kappa$ B ligand (RANKL) secretion by bone marrow stem cells (BMSCs), which attract circulating tumour cells (part e). Stress factors promote the release of various pro-angiogenic (part f) and inflammatory (part g) factors, such as vascular endothelial growth factor (VEGF), interleukin-8 (IL-8), IL-6 and prostaglandins, from tumour and stromal cells, all of which promote tumour progression. Stress-induced immune suppression facilitates tumour immune evasion by upregulation of myeloid-derived suppressor cells (MDSCs), regulatory T cells and M2 tumour-associated macrophage (TAM) polarization, and through downregulation of effector T cell and natural killer cell activity (part h). Activation of prostaglandin receptors and activation of  $\beta$ -ARs each induces the same intracellular downstream processes (not shown), including the cyclic AMP (cAMP)–protein kinase A (PKA) pathway, suggesting that simultaneous blockade of  $\beta$ -adrenergic and prostanoid signalling might be important to improve cancer treatment.

Table 1 | **Biological effects of stress on cancer progression: preclinical studies and related observations in patients with cancer**

Cancer	Model	Stressor	Effect (location)	Mediator	Refs
<b>Angiogenesis</b>					
Melanoma, breast, ovarian	Human cells in vitro	Adrenaline or noradrenaline	↑ Angiogenesis; ↑ VEGF; ↑ IL-6; ↑ IL-8	Tumour–endothelial cell contact ( $\beta_2$ -AR–Jagged 1–Notch); tumour cell $\beta_1$ -AR and/or $\beta_2$ -AR–cAMP–PKA signalling	81,95–98
Ovarian <sup>a</sup> , pancreatic <sup>a</sup> , colorectal <sup>b</sup> , mammary <sup>b</sup> , prostate <sup>a</sup>	Mice; human or mouse cells in vitro	Social isolation, chronic restraint, audio of screaming rats, laparotomy or orisoproterenol	↑ Tumour vascularization; ↑ tumour VEGF; ↑ tumour growth; ↓ TSP1	$\beta_2$ -AR–cAMP–PKA signalling; ↑ HIF1 $\alpha$ ; ↓ CXCL4; macrophage recruitment; $\beta$ -AR–CREB–HDAC2 pathway	41,87, 99–102
Ovarian	Patients with cancer <sup>c</sup>	Low social support <sup>d</sup> or helplessness	↑ Plasma VEGF; ↑ tumour VEGF	NA	103,104
Ovarian	Patients with cancer <sup>c</sup>	Low social attachment <sup>d</sup> or vegetative depression <sup>d</sup>	↑ IL-6 (plasma, ascites); ↑ nocturnal cortisol (saliva)	NA	105,106, 220
<b>Lymphatic modulation</b>					
Breast <sup>a,b</sup>	Mice	Chronic restraint	↑ Tumour VEGFC; ↑ tumour LVD; ↑ lymphatic dilation, flow; ↑ lymph node metastasis	$\beta$ -AR; ↑ COX2; macrophage recruitment	108
Breast <sup>c</sup> , cervical	Patients with cancer	Social isolation <sup>d</sup> or SNS activity	↑ Tumour LVD; ↑ lymphatic flow	NA	108,109
<b>Inflammation and immunity</b>					
Mammary <sup>b</sup> , leukaemia <sup>b</sup>	Rats; blood samples from stressed rats studied ex vivo	Laparotomy, swim stress, wet cage, metaproterenol or adrenaline	↓ NKCC	$\beta_1$ -AR and/or $\beta_2$ -AR	33,40, 112–114
Colorectal <sup>b</sup> , squamous cell carcinoma <sup>b</sup> , mammary <sup>b</sup> or melanoma <sup>b</sup>	Mice; mouse cells in vitro	Chronic restraint, 22 °C housing temperature, audio of screaming mice or laparotomy	↓ T <sub>H</sub> 1 cell/T <sub>H</sub> 2 cell-type cytokine ratio (serum); ↓ effector CD8 <sup>+</sup> and CD4 <sup>+</sup> TILs; ↑ tumour growth; ↑ MDSCs (tumour, spleen); ↑ regulatory T cells (tumour, blood)	↓ CXCL4; $\beta$ -AR; $\beta_2$ -AR–STAT3 signalling	62,84,100, 116,118
Ovarian or breast	Patients with cancer <sup>c</sup>	Low social support <sup>d</sup> , high distress <sup>d</sup> , depressed/anxious mood <sup>d</sup> or psychological stress	↓ NKCC (tumour, blood); ↓ T <sub>H</sub> 1 cell/T <sub>H</sub> 2 cell-type cytokine ratio (blood, ascites, tumour); ↓ MDSCs (blood)	NA	115,117,119
Breast <sup>a,b</sup> or ovarian <sup>a</sup>	Mice; human or mouse cells in vitro	Chronic restraint or social isolation	↑ Macrophage recruitment; ↑ prostaglandin (tumour cells, TAMs); ↑ TAM M2 polarization	$\beta$ -AR; $\beta_2$ -AR/NF- $\kappa$ B–prostaglandin E <sub>2</sub> axis; $\beta$ -AR–cAMP–PKA–MCP1 production	41,79,90, 108,120
Ovarian or breast	Patients with cancer <sup>c</sup>	Psychological stress, depression or social isolation <sup>d</sup>	↑ Plasma IL-1 $\alpha$ ; ↑ tumour prostaglandin; ↑ M2 polarization of TAMs	NA	79,109,119
<b>Metastasis</b>					
Breast <sup>a,b</sup> , gastric <sup>a</sup> or pancreatic <sup>a</sup>	Mice; human or mouse cells in vitro	Chronic restraint, alternating stressors, or audio of screaming rats	↑ Pre-metastatic niche; ↑ EMT; ↑ MMPs (tumour, stroma)	$\beta$ -AR–RANKL; $\beta$ -AR–CCL2/CCR2 axis; miR-337-3p–STAT3	41,42,74, 80,82,85, 99,122
Breast <sup>a,b</sup> or gastric <sup>a</sup>	Mice or rats	Chronic restraint, laparotomy, alternating stressors, wet cage or swim stress	↑ Spontaneous and experimental metastasis	$\beta_1$ -AR and/or $\beta_2$ -AR	33,40,41, 74,85,108, 114,122,125
Ovarian or breast	Patients with cancer <sup>c</sup>	Perceived stress <sup>d</sup> , social isolation <sup>d</sup> or depression <sup>d</sup>	↑ Tumour EMT genes; ↑ TAM MMP9	NA	104,109,124

All findings are causal, except those indicated as correlational findings in the 'Model' or 'Cancer' column. ↑, increase; ↓, decrease;  $\beta$ -AR,  $\beta$ -adrenergic receptor; cAMP, cyclic AMP; CCL2, CC-chemokine ligand 2; CCR2, CC-chemokine receptor 2; COX2, cyclooxygenase 2; EMT, epithelial–mesenchymal transition; HDAC2, histone deacetylase 2; IL-6, interleukin-6; LVD, lymphatic vessel density; MDSC, myeloid-derived suppressor cell; MMP, matrix metalloproteinase; NA, not applicable; NKCC, natural killer cell cytotoxicity; NF- $\kappa$ B, nuclear factor- $\kappa$ B; PKA, protein kinase A; RANKL, receptor activator of nuclear factor- $\kappa$ B ligand; SNS, sympathetic nervous system; TAM, tumour-associated macrophage; T<sub>H</sub>1 cell, T helper 1 cell; T<sub>H</sub>2 cell, T helper 2 cell; TIL, tumour-infiltrating lymphocyte; TSP1, thrombospondin 1; VEGF, vascular endothelial growth factor. <sup>a</sup>Xenograft. <sup>b</sup>Syngeneic. <sup>c</sup>Correlational findings. <sup>d</sup>Adjusted for disease stage.

null effect, whereas 18% indicated harmful effects and 6% indicated protective effects.

More recent studies linked various specific stressors, including a cold climate<sup>148</sup>, bereavement<sup>61</sup>, war<sup>149</sup> and depression<sup>150</sup>, to higher incidence of various cancer types,

yet other studies reported null effects<sup>151–153</sup>. Focusing on work stress as a risk factor, two meta-analyses yielded inconsistent conclusions: the first<sup>154</sup> reported null effects of prospective studies, whereas the second<sup>155</sup> reported elevated relative risk (of 1.24 and 1.36 in lung cancer



and CRC, respectively), but the latter also included case-control studies that are susceptible to retrospective recall and interpretation bias. Last, it is important to note that in humans, malignant transformation is a prolonged process and subclinical cancer dormancy is highly prevalent<sup>30</sup>. Thus, cancer incidence may be elevated not only by initiation of the disease but also by escape from dormancy or faster progression of cancer to clinical manifestation. Indeed, animal studies report that stress and stress factors can induce escape from dormancy in tumour cells<sup>156–158</sup>.

#### Laparotomy

An experimental stress paradigm in which a midline abdominal incision is performed under anaesthesia, and often the small intestine is externalized and left hydrated in a soaked gauze pad for 30 min. The intestine is then internalized and the abdomen is sutured.

#### Social confrontation stress

An experimental stress paradigm where an intruder rodent (a non-cage-mate animal) is introduced into a home cage populated with several stable cage-mates. The intruder is usually attacked by the residents cage-mates and/or displays submissive behaviour.

#### Foot shock stress

An experimental stress paradigm that is executed in an apparatus containing an electrified grid floor, in which the animal is exposed to electric shocks of varying intensity and duration. The paradigm can be acute or chronic, and is also used for fear-conditioning.

#### Hazard ratio

The ratio of the probability of events in a treatment group to the probability of events in a control group.

#### Publication bias

The tendency to publish a study based on its results (positive rather than negative findings or significant rather than non-significant findings). Existence of this bias can be statistically assessed in meta-analyses by Egger's linear regression test.

#### Cochrane

A non-profit organization (maintaining no conflict of interests), which, among other activities, publishes methodologies and guidelines to produce high-quality systematic reviews and meta-analyses.

### Stress and cancer progression

Effects of stress on cancer progression are commonly studied by assessing survival rates in patients diagnosed with cancer. The overall hazard ratio indicated by 157 prospective studies included in the 2008 meta-analysis discussed above<sup>146</sup> was 1.03 (95% CI 1.02–1.04,  $P < 0.001$ ), with more than 73% of studies reporting null findings. This small effect should be interpreted with caution. First, stress (for example, life events) was commonly assessed irrespective of its timing relative to cancer detection, and the specific impact of stress while having cancer, including the critical perioperative period, was not assessed. Second, most patients with cancer experience some levels of cancer-related distress<sup>159,160</sup>, which may suffice to generate a similar effect on cancer progression, irrespective of whether patients were categorized with low versus high stress levels. This could mask relations between stress levels and cancer progression in such circumstances, but nevertheless could enable marked beneficial effects of stress-reducing interventions. Third, although comprehensive, this meta-analysis is 13 years old, and has narrowed down analyses to either distinct cancer types or defined stressors. More recent meta-analyses have focused on more specific conditions, and have reported larger effect sizes. Specifically, depression in patients with breast cancer predicted 29% elevated risk for cancer-specific mortality<sup>161</sup>, and low levels of perceived social support, a smaller social network, being unmarried or being depressed predicted a 12–25% elevated relative risk for cancer mortality in various cancer types<sup>162,163</sup>.

Indeed, recent studies confirmed that the effects of stress on survival are stressor-specific and cancer-specific. For example, depression that followed cancer diagnosis predicted decreased survival in breast<sup>161</sup> and renal<sup>164</sup> cancers, but not in ovarian cancer<sup>165</sup>. Low social support and low social attachment predicted decreased survival in patients with ovarian cancer<sup>165</sup>, breast cancer<sup>166</sup> or CRC<sup>167</sup>, whereas work stress had no effect<sup>152</sup>. Importantly, previous life history of stress and adversities may interact with post-diagnosis stress<sup>168,169</sup>, as early adverse experiences can shape maladaptive responses to stressors<sup>27</sup>.

Overall, given the small and inconsistent effects reported by epidemiological studies, and the heterogeneous methodological approaches, populations studied and type of stressors, it remains uncertain whether stress can increase cancer incidence, and to what extent it facilitates cancer progression. Potentially, stress has a larger impact in certain conditions or populations. Clearly,

epidemiological studies face significant obstacles. The subjective perception of stress in patients with cancer is influenced by the physical and mental burden of the disease, and therefore studies that retrospectively assess pre-diagnostic or post-diagnostic stress by subjective reports are biased<sup>147</sup>. On the other hand, objective exposure to adverse life events (for example, based on national registries of divorces or deaths) does not include the individual subjective experience. As described below, the use of stress-reducing interventions in RCTs can circumvent many of these obstacles.

### Stress management in patients with cancer

The most methodologically sound approach to test in humans whether stress affects cancer progression would be RCTs, where the intervention is a verified stress-management approach and the outcomes include psychological indices, interim biomarkers and, most importantly, long-term cancer outcomes. Such RCTs are not practical for studying cancer incidence but are feasible for studying cancer progression and mortality. Such psychological and pharmacological RCTs have been conducted during the last four decades, as discussed below.

#### Psychological RCTs: long-term outcomes

Recent meta-analyses<sup>170–173</sup> have cumulatively identified 22 RCTs that employed psychosocial interventions as being methodologically stringent, using Cochrane or other criteria. Most interventions were initiated at least a month postoperatively (16/22 RCTs) and/or were conducted in patients with metastatic disease (12/22 RCTs); and most studies employed group interventions (14/22 RCTs), rather than individual (7/22 RCTs) approaches. Importantly, most interventions did yield improvement in psychological indices (TABLE 2), and a few improved physiological biomarkers (for example, natural killer cell activity)<sup>174–176</sup> (BOX 4). Based on these meta-analyses (each considering 11–15 trials)<sup>170–173</sup> and our own assessment of all 22 studies (TABLE 2), there is no clear evidence for improvements in long-term cancer outcomes<sup>171,172</sup>, but the results are nevertheless informative. Specifically, there seems to be an agreement that some interventions can delay disease progression during the first post-intervention years, but less so or not at all beyond this initial period<sup>171,172</sup>. Psychosocial interventions may have temporary effects either because their impact on tumour biology is short-lasting or because patients' adherence to the psychological intervention reduces along the follow-up period. It is suggested that some patients, more than others, may benefit from psychological interventions, specifically patients who are older, unmarried and psychologically vulnerable or stressed<sup>8,170</sup>, as well as patients in earlier disease stages (for example, early-stage melanoma)<sup>177</sup>. It should be noted that some of these studies have been criticized for having methodological flaws<sup>178–180</sup> (but also see the response to criticism)<sup>181</sup>, including not having the statistical power to study cancer mortality, employing only 30–150 patients per group, which may lead to exaggerated effect sizes. Some interventions have been suggested to act through improving patients' treatment

Table 2 | Psychosocial stress-reducing interventions in RCTs and long-term cancer outcomes

Study	Patient numbers	Intervention (setting; timing; duration (weeks) <sup>a</sup> ; treatment type)	Psychological benefit	Survival effect	Survival effect size <sup>b</sup>
<b>Early-stage breast cancer</b>					
Burton et al. (1995) <sup>187</sup>	n = 200, 4 groups of 50 each <sup>c</sup>	Individual; preoperative; 1; one interview + 30-min psychotherapeutic intervention	Yes	No	First-year recurrence rates: T = 7–10%; C = 14%; simple contrast between control and intervention groups; NS
Kissane et al. (2004) <sup>257</sup>	n = 303, T = 154	Group; post surgery; 20; CBT-supportive therapy sessions + 3 relaxation sessions	Yes	No	Median survival time (months): T = 81.9, C = 85.5; multivariate Cox analysis, HR = 1.35, NS
Andersen et al. (2008) <sup>258</sup>	n = 227, T = 114	Group; post surgery; 16 weekly sessions + 8 monthly sessions; stress management	Yes	Yes	Mortality, 11-year follow-up: T = 24/114, C = 30/113; multivariate Cox analysis, HR = 0.44; P = 0.016 Median time to recurrence (months): T = 33.6, C = 26.4; multivariate Cox analysis, HR = 0.55, P = 0.034
Boesen et al. (2011) <sup>259</sup>	n = 210, T = 105	Group; post surgery; 8; comprehensive psychoeducation + supportive therapy	No	No	Mortality, 4-year follow-up: T = 6/105, C = 3/105; statistical analysis not preformed due to low event number
Stagl et al. (2015) <sup>260</sup>	n = 240, T = 120	Group; post surgery; 10; cognitive-based stress management	Yes	Yes <sup>d</sup>	Mortality, 8–15-year follow-up: T = 15/120, C = 15/120; multivariate Cox analysis using four covariates <sup>d</sup> , HR = 0.21, P = 0.04
<b>Metastatic breast cancer</b>					
Spiegel et al. (1989) <sup>182</sup>	n = 86, T = 50	Group; post surgery; 52; supportive-expressive therapy + self-hypnosis	Yes	Yes	Mean survival time (months): T = 36.6, C = 18.9; log-rank test, P < 0.0001
Cunningham et al. (1998) <sup>261</sup>	n = 66, T = 30	Group; post surgery; 35; supportive + CBT	No	No	Median survival time (months): T = 28.8, C = 23.6; log-rank test, P = 0.35
Edelman et al. (1999) <sup>262</sup>	n = 124, T = 62	Group; post surgery; 8 weekly sessions + 3 sessions once a month; CBT	Yes <sup>e</sup>	No	Median survival time (months): T = 11.64, C = 12.84; log-rank test, NS
Goodwin et al. (2001) <sup>184</sup>	n = 225, T = 158	Group; replication study, similar <sup>f</sup> to Spiegel et al. (1989) <sup>182</sup>	Yes <sup>g</sup>	No	Median survival time (months): T = 17.9, C = 17.6; Cox univariate analysis, HR = 1.06, NS
Kissane et al. (2007) <sup>263</sup>	n = 227, T = 147	Group; similar to Spiegel et al. (1989) <sup>182</sup> + 3 relaxation classes	Yes	No	Median Survival time (months): T = 24, C = 18.3; univariate Cox analysis, HR = 0.92, NS
Spiegel et al. (2007) <sup>183</sup>	n = 125, T = 64	Group; replication study, same as Spiegel et al. (1989) <sup>182</sup>	Yes	No/yes <sup>h</sup>	Median survival time (months): exploratory subgroup findings (n = 25 ER-negative <sup>h</sup> ); T = 29.8, C = 9.3; Multivariate Cox analysis, P = 0.002
Andersen et al. (2010) <sup>264</sup>	n = 62, T = 29 (a subgroup of patients from Andersen et al. (2008) <sup>258</sup> ) <sup>i</sup>	Group; same as Andersen et al. (2008) <sup>258</sup>	Yes	Yes	Mortality after recurrence: T = 19/29, C = 25/33; median survival after recurrence (months): T = 38.4, C = 20.4; multivariate Cox analysis, HR = 0.41, P = 0.014
<b>Melanoma</b>					
Fawzy and Fawzy (2003) <sup>177</sup>	n = 68, T = 34	Group; post surgery; 6; health education + stress management + coping skills + psychological support	Yes	No	Mortality, 5–6-year follow-up: T = 3/34, C = 10/34; log-rank test, P = 0.03 Mortality, 10-year follow-up: T = 9/34, C = 11/34; log-rank test, NS
Boesen et al. (2007) <sup>185</sup>	n = 262, T = 131	Group; replication study, similar to Fawzy and Fawzy (2003) <sup>177</sup>	Yes <sup>j</sup>	No	Mortality, 4–6-year follow-up: T = 8/128, C = 8/130; univariate Cox analysis, HR = 0.99, NS
<b>Other cancer types</b>					
Linn et al. (1982) <sup>265</sup> (several cancer types)	n = 120, T = 62	Individual; NR; NR; supportive therapy	Yes	No	Mean survival time (months), 1-year follow-up: T = 3.7, C = 4.37; life table method, $\chi^2$ test, NS
Illychkyj et al. (1994) <sup>266</sup> (several cancer types)	n = 127, four groups: <sup>k</sup> T = 31, 30, 35, C = 31	Group; NR; 24; supportive discussion group sessions	No	No	Mean survival time (months), 10-year follow-up: T = 70.7, C = 82.4; log-rank test, NS

Table 2 (cont.) | Psychosocial stress-reducing interventions in RCTs and long-term cancer outcomes

Study	Patient numbers	Intervention (setting; timing; duration (weeks) <sup>a</sup> ; treatment type)	Psychological benefit	Survival effect	Survival effect size <sup>b</sup>
<b>Other cancer types (cont.)</b>					
Ratcliffe et al. (1995) <sup>267</sup> (lymphoma)	n = 63, T = 36	Individual; post third cycle of chemotherapy; NR; relaxation training with or without hypnosis	Yes	Yes	Mortality, 5-year follow-up: T = 14/36, C = 13/27; multivariate Cox analysis, HR = 0.66, P = 0.06
Kuchler et al. (2007) <sup>188</sup> (gastrointestinal cancers)	n = 271, T = 136	Individual; pre surgery to discharge from hospital; 2–25 sessions; individually tailored psychological support	Yes	Yes	Survival, 2-year follow-up: T = 69/136, C = 45/135; log-rank test, P = 0.002; survival, 10-year follow-up: T = 29/136, C = 13/135; log-rank test, P = 0.006
Ross et al. (2009) <sup>268</sup> (colorectal cancer)	n = 249, T = 125	Individual; post surgery; 10 meetings over 24 months; home visits by a medical doctor or nurse providing emotional support or information	No	No	Mortality, 6.5–9.5-year follow-up: T = 75/125, C = 73/124; log-rank test, NS
Temel et al. (2010) <sup>269</sup> (metastatic non-small cell lung cancer)	n = 151, T = 77	NR; intervention group patients were assigned to early palliative care <sup>l</sup>	Yes	Yes <sup>l</sup>	Median survival time (months): T = 11.6, C = 8.9; log-rank test, P = 0.02
Guo et al. (2013) <sup>270</sup> (several cancer types)	n = 178, T = 89	Individual; during radiotherapy; 4–6; psychoeducation + CBT + supportive-expressive therapy	Yes	No	% survival, 2-year follow-up: T = 83.1%, C = 84.3%; log-rank test, NS
Zhang et al. (2013) <sup>189</sup> (oesophageal cancer)	n = 60, T = 31	Individual; pre surgery; 3 weeks, sessions every other day; health education, psychological support, stress management, coping strategies and behaviour training	Yes	No	Survival, 4-year follow-up: T = 15/27, C = 18/28; log-rank test, NS

C, control group; CBT, cognitive behavioural therapy; ER, oestrogen receptor; NR, not reported; NS, not significant; RCT, randomized controlled trial; T, treatment group. <sup>a</sup>One weekly session, unless otherwise specified. <sup>b</sup>Log-rank test and univariate Cox analyses address differences between groups that are driven only by group assignment, whereas multivariate Cox analyses incorporate additional factors into the statistical model beyond group assignment. <sup>c</sup>The different groups were: preoperative interview; preoperative interview + 30-min preoperative psychotherapeutic intervention; preoperative interview + chat (attention); and routine hospital care control. <sup>d</sup>Equivalent number of deaths between groups; difference was statistically significant in a Cox multivariate analysis addressing age at diagnosis, disease stage, tumour size, HER2 status and hormonal treatment. <sup>e</sup>Improved psychological measures at the end of the intervention were not sustained at 3 and 6-month follow-up. <sup>f</sup>Relaxation techniques were taught instead of self-hypnosis. <sup>g</sup>In patients with high baseline of distress. <sup>h</sup>Cox proportional hazard analysis showed a significant interaction between ER status and treatment, indicating that ER-negative patients allocated to the intervention survived longer than control patients. <sup>i</sup>Participants in this study were patients who previously participated in Andersen et al. study<sup>258</sup>. <sup>j</sup>Psychological benefits were only evident shortly after the intervention, and enrolled patients exhibited low baseline levels of psychological distress. <sup>k</sup>The different groups were: group meetings professionally guided by a social worker for 6 months; group meetings professionally guided for 3 months + 3 months of unguided meetings; unguided group meetings; and control (no group meetings). <sup>l</sup>Earlier initiation of palliative care, also addressing individual psychosocial needs of the patients.

adherence, patients' health behaviour and quality of the medical treatment (for example, additional surveillance and care) following improved communication with medical personnel<sup>180</sup>. Thus far, no research group has replicated previously reported positive outcomes, although given the objective difficulties of intervention trials and lack of funding, only a few replications have been attempted<sup>177,182–185</sup>. Notably, each of the 22 studies used a different treatment protocol, initiated treatment at different times during cancer progression, provided treatment for a different duration and/or studied a different patient population and cancer type (TABLE 2). These heterogeneities may be the source of inconsistent outcomes, and different results of meta-analyses. At the single study level, 8 of the 22 interventions reported a significant survival advantage of a psychosocial intervention (TABLE 2). Beyond the legitimate debate of the validity of specific studies, eight successful demonstrations could indicate promising outcomes. However, as only eight such demonstrations have been reported, and the results of these eight interventions have not been replicated in published studies, combined with the likelihood of unpublished studies with null effects, this raises questions regarding the effectiveness of these psychosocial interventions in improving cancer survival.

However, we believe that these inconsistent outcomes are expected a priori, given the following considerations. First, as discussed above and in BOX 3, critical time periods, such as the immediate perioperative time frame in patients undergoing surgery, may bear a non-proportional high impact on the fate of metastatic disease, especially in patients harbouring only scattered tumour cells and micrometastases<sup>186</sup>. Psychological interventions have been commonly initiated weeks following surgery, which would miss this critical period (only 3/22 studies in TABLE 2 are perioperative)<sup>187–189</sup>. Such delayed interventions may impact metastases at a more advanced and therapeutically resistant stage, thus confronting a greater challenge in preventing metastatic disease, but still having the ability to delay a metastatic outbreak<sup>171,172,174,177</sup>. Second, many medical procedures, including surgery and chemotherapy, induce stress-related inflammatory responses of local physiological origin, including cellular responses of injured tissue (for example, increased levels of damage-associated molecular patterns and prostaglandins) (BOX 3). Psychosocial interventions alone are unlikely to significantly reduce such local responses, which may mask the potential beneficial effects of psychosocial interventions during medical procedures (BOX 4). Third, in many

**Box 4 | Behavioural stress management and its impact on short-term cancer-related indices**

Multiple psychological, behavioural and physiological interventions have been used to target different aspects of stress in patients with cancer, such as massage, acupuncture, yoga, tai chi, mindfulness and cognitive behavioural stress-reduction interventions (reviewed in REFS<sup>8,10,12</sup>). Such interventions were shown to reduce stress, anxiety and depression, and to improve quality of life<sup>309,310</sup> in patients with cancer (for example, in breast cancer<sup>174</sup> and melanoma<sup>175</sup>). Accordingly, current guidelines for optimal oncological care include screening and addressing psychosocial concerns<sup>311</sup>.

Importantly, Antoni and Dhabhar<sup>8</sup> suggested that stress-management interventions can have physiological protective effects against tumour progression through improving protective immunity (for example, immunosurveillance), reducing chronic inflammatory processes and inhibiting immunosuppressive mechanisms (for example, regulatory T cell activity). Indeed, in breast cancer survivors, yoga and tai chi reduced pro-inflammatory processes<sup>312,313</sup>, and mindfulness-based stress reduction increased the T helper 1 cell (T<sub>H</sub>1)/T helper 2 cell (T<sub>H</sub>2) ratio<sup>314</sup>, decreased nuclear factor- $\kappa$ B (NF- $\kappa$ B) activity and increased anti-inflammatory signalling and gene expression of type 1 interferon<sup>315</sup>. Similar effects were noted by Antoni, studying the effects of a cognitive behavioural therapy (CBT)-based stress-management intervention in patients with breast cancer following surgery<sup>316</sup>. In addition to significant psychological benefits, the intervention enhanced protective immunity (that is, increased gene expression of type 1 interferon, and serum levels of interferon- $\gamma$  (IFN $\gamma$ ) and interleukin-2 (IL-2)), and reduced inflammatory processes (for example, reduced expression of the genes encoding IL-1 $\beta$ , IL-6 and TNF, and increased prevalence of glucocorticoid receptor (GR) response elements)<sup>176,317</sup>.

Missing from these studies are specific assessments of sympathetic activity and potential reduction of tumour-associated noradrenaline and/or systemic adrenaline levels in treated patients with cancer. Correlative studies in patients with cancer do suggest association of these indices with stressors such as social isolation<sup>91</sup>.

Taken together, these changes may predict favourable prognosis for a broad range of patients with cancer, and were suggested by Antoni and Dhabhar<sup>8</sup> to explain the beneficial effects of stress-management interventions on long-term cancer survival<sup>258,260,264</sup>. Such interventions should be initiated as early as possible after cancer diagnosis, and potentially before cancer surgery<sup>197</sup>, to improve their impact on both mental health and long-term cancer outcomes.

patients, psychosocial interventions cannot be expected to be effective, either given low stress levels at study entry or given individual characteristics of psychological needs or coping style, not addressed by prevalent standardized group therapies. Last, if one expects the effect size of psychological intervention to be similar to those of chemotherapy or hormonal therapy, hundreds of patients of the same cancer type would need to be included. We assert that, given appropriate funding, all of these obstacles can be overcome, as detailed below, enabling better assessment of the efficacy of stress management for improving cancer survival.

**Pharmacological RCTs: cancer biomarkers**

Recently, several biomarker RCTs have employed pharmacological interventions to antagonize stress responses in patients with cancer, all employing the non-selective  $\beta$ -blocker propranolol. Among other reasons, this drug was chosen based on its early promising outcomes in animal models of stress or surgery-induced cancer progression<sup>41,74,85,108,125</sup>, the involvement of both  $\beta_1$ -AR and  $\beta_2$ -AR in various pro-malignant mechanisms<sup>6,40,65,87</sup> and its high safety profile relative to other adrenergic antagonists, especially regarding potential cardiovascular and tissue healing-related complications<sup>190–192</sup>. Among other positive prognostic outcomes in treated patients, propranolol downregulated the expression of mesenchymal

genes, the EMT transcription factors Snail and Slug, and activity levels of the inflammatory transcription factors nuclear factor- $\kappa$ B (NF- $\kappa$ B) and AP-1 in primary breast tumours<sup>193</sup>, facilitated a decrease in CA-125 serum levels in ovarian cancer<sup>194</sup> and decreased classical monocyte activation in haematopoietic cell transplant recipients<sup>195</sup>. Propranolol is also currently being tested in combination with immunotherapy in patients with melanoma<sup>196</sup>.

It is important to note that adrenergic stress responses and inflammatory responses often intertwine, especially during cancer treatments, as perioperative stress, tissue damage and other medical procedures simultaneously induce both adrenergic and prostanoid responses<sup>197–199</sup> (BOX 3), because each response facilitates the other<sup>197</sup> and because  $\beta$ -adrenergic and prostaglandin receptors activate the same intracellular immunosuppressive and tumour-promoting mechanisms (for example, cAMP-PKA signalling)<sup>197</sup>. Therefore, it may be necessary to simultaneously block  $\beta$ -adrenergic and inflammatory responses to overcome the metastatic promoting effects of stress and/or medical procedures. Indeed, several preclinical studies indicated that simultaneous blockade of  $\beta$ -AR and COX2 activity (using propranolol and etodolac, respectively) was synergistically more effective than each approach alone in preventing immunosuppression and cancer metastasis<sup>33,125,200,201</sup>.

These insights have been recently implemented clinically in the context of curative oncological surgeries, in two RCTs that have initiated combined propranolol and etodolac treatment 5 days before surgery, for a total of 11–20 days, in patients with breast cancer<sup>190,202</sup> or CRC<sup>203</sup>. In resected tumours from both RCTs, the treatment decreased EMT and the activity of several pro-metastatic and pro-inflammatory transcription factors (for example, those of the GATA, STAT, EGR and CREB families), and improved the profile of infiltrating leukocytes and tumour proliferation markers (for example, Ki-67)<sup>190,202,203</sup>. In patients with breast cancer, where repeated perioperative blood samples were also analysed, treatment improved systemic inflammatory and immunological markers, including IL-6, C-reactive protein (CRP) and natural killer cell CD11a expression, before and/or after surgery<sup>190,202</sup>. Although not powered to assess survival, the treatment improved 3-year disease-free survival (DFS) in patients with CRC who were protocol compliant<sup>203</sup>, and our as yet unpublished data also show improved 5-year DFS.

Overall, these clinical findings indicate that  $\beta$ -adrenergic blockade, with or without COX2 inhibition, can significantly improve numerous biomarkers of cancer progression, and justify larger RCTs to test long-term cancer outcomes of pharmacological stress management, as currently being conducted (NCT03838029 (REF.204), NCT03919461 (REF.205)).

Additional pharmacological approaches were also studied. Specifically, the use of anxiolytic and antidepressant drugs (for example, selective serotonin reuptake inhibitors and serotonin and noradrenaline reuptake inhibitors) in patients with cancer is prevalent and effective in reducing anxiety and depression<sup>10,16</sup>. Nevertheless, epidemiological studies assessing their impact on cancer



### CpG class C

(CpG-C). A synthetic oligodeoxynucleotide (ODN) that functions as a Toll-like receptor 9 (TLR9) agonist and induces a physiological host-dependent activation of the immune system.

### Glucopyranosyl lipid-A stable emulsion

(GLA-SE). A synthetic agonist of Toll-like receptor 4 (TLR4). For administration, GLA is dissolved in an oil–water stable emulsion that serves as an adjuvant delivery system.

survival yielded inconsistent results<sup>206–208</sup>, and no effects on cancer survivorship were noted when causally assessed in an RCT that enrolled patients with advanced cancer of various types<sup>209</sup>. Additionally, the effects of anxiolytic and antidepressant drugs on cancer-related biomarkers is largely unknown, and their impact on such indices in controlled preclinical studies is contradictory<sup>210–212</sup>. Thus, more preclinical and clinical research is needed to assess the impact of such pharmacological approaches on cancer-related biomarkers and long-term outcomes.

### Stress and cancer reciprocal relations

In the clinical setting, stress and cancer can promote each other. Patients with cancer often experience peaks of stress on initial diagnosis, on cancer treatment and on cancer recurrence<sup>159,160,198,213,214</sup>. Throughout cancer survivorship, anxiety decreases in some patients but persists in others<sup>169</sup>, and patients with cancer show increased risk for anxiety and depressive disorders<sup>214–216</sup>. Consequently, stress responses and affective disorders may accelerate cancer progression through various mechanisms detailed above. Indeed, among patients with breast cancer, higher anxiety, stress, depressive symptoms or elevated diurnal cortisol levels were found to predict suppressed antitumour cell-mediated immunity<sup>217–219</sup>; and perceived stress, social isolation and depression predicted increased tumour cell EMT and levels of MMPs in patients with ovarian and breast cancer (controlling for disease parameters) (TABLE 1).

Simultaneously, the malignant tissue itself may heighten local and systemic stress responses, through tumour-induced increases in sympathetic tumour innervation and noradrenaline release<sup>65</sup>, and through local and systemic inflammation that affects the CNS, dysregulates HPA axis activity<sup>220,221</sup> and facilitates depression, sleep disturbances and cancer-related fatigue<sup>222–224</sup>. Together with cancer-related cognitive impairments<sup>225,226</sup>, these symptoms may induce or exacerbate stress responses<sup>227</sup>, perpetuating a vicious cycle of stress and cancer (FIG. 1).

Importantly, the brain, tumours and the immune system all affect each other bidirectionally, either promoting or hindering tumour progression. For example, artificial activation of the brain reward system in mice was found to decrease a suppressive MDSC phenotype through reduced SNS signalling, resulting in attenuated tumour growth<sup>228</sup>. Crosstalk between stress and cancer is prominent within the perioperative period. In patients with breast, colorectal or ovarian cancer, plasma cortisol levels and/or stress inflammatory indices were elevated even before surgery, presumably due to psychological distress or tumour-derived inflammation<sup>190,203,220</sup>, which may sensitize pain responses and worsen psychological stress<sup>197</sup>. Pharmacological blockade of stress and/or inflammatory responses before surgery reduces these indices, as well as tumour EMT and other pro-metastatic molecular indices in the malignant tissue<sup>190,202,203</sup>.

### Stress impairs cancer treatments

Stress was reported in both preclinical and clinical studies to impair adjuvant and neoadjuvant cancer treatments, including chemotherapy, radiotherapy and

immunotherapy, through mediation of glucocorticoids and/or catecholamines. Specifically, in murine models, behavioural and/or surgical stress impaired the capacity of the (clinically studied)<sup>229</sup> immunostimulating agents, CpG class C (CpG-C) and glucopyranosyl lipid-A stable emulsion (GLA-SE), to reduce experimental metastases in mammary cancer and CRC models<sup>133,230,231</sup>; and in vitro, corticosterone suppressed IL-12 secretion from leukocytes following CpG-C or GLA-SE stimulation<sup>133,232</sup>. Social disruption stress or  $\beta$ -AR activation in melanoma and lymphoma mouse models compromised several immunotherapies through impairing CD8<sup>+</sup> T cell responses<sup>233,234</sup>; and restraint stress, catecholamines or glucocorticoids impaired the efficacy of chemotherapy in human breast and ovarian cancer cell lines, both in vitro and in xenograft models<sup>52,235</sup>.

Additionally, treatment with cytotoxic therapy or sunitinib (an inhibitor of several tyrosine-kinase receptors exerting both anti-angiogenic and direct anti-tumour effects) was impaired by chronic restraint stress or administration of noradrenaline or adrenaline in CRC, prostate cancer and melanoma mouse models<sup>236–238</sup>. In mammary, pancreatic, melanoma, colon and lung cancer models,  $\beta$ -AR signalling, induced by ambient temperature stress, jeopardized cytotoxic therapies (cisplatin and nab-paclitaxel chemotherapies and TRAIL (TNF-related cytokine which induces apoptosis by binding to cell surface death receptors))<sup>239</sup>, radiotherapy<sup>240</sup> and PD1-targeted immunotherapy<sup>84</sup>. Activation of  $\beta$ -AR also induced resistance to the HER2 targeted therapy trastuzumab in gastric and breast cancer mouse models<sup>241,242</sup>. Social disruption and acute restraint stress impaired chemotherapy and immunotherapy in lung cancer, CRC and fibrosarcoma mouse models, through glucocorticoid-induced expression of the immunosuppressive transcription factor TSC22D3 in dendritic cells, and consequent impairment of anti-tumour immunity<sup>92</sup>. Administration of the synthetic glucocorticoid dexamethasone induced chemotherapy and hormone-therapy resistance in prostate and breast cancer mouse models<sup>93,243–245</sup>, as well as in vitro in breast cancer tumour samples and numerous human carcinoma cell lines<sup>243,246</sup>. Last, in mice, blockade of GR in combination with chemotherapy or hormone therapy potentiated in vivo therapeutic responses<sup>244,245</sup>.

Corresponding clinical observations have been reported in patients. In breast cancer, tumour expression of  $\beta$ -AR negatively correlated with response to trastuzumab<sup>242</sup>, and in patients with prostate cancer, increased GR expression in bone metastases following treatment with enzalutamide (anti-androgen receptor therapy) predicted poorer therapeutic response<sup>244</sup>. Retrospective observations indicated that incidental  $\beta$ -blocker usage with anti-angiogenic agents, immunotherapy, radiation and/or chemotherapy extended patient DFS and overall survival<sup>247–250</sup>.

In sum, ample preclinical studies indicate that stress, noradrenaline, adrenaline and glucocorticoids can jeopardize adjuvant and neoadjuvant therapies, although clinical studies have not sufficiently addressed this important issue. Also concerning is the prevalent use of synthetic glucocorticoids (including dexamethasone) in

patients with cancer. These agents are routinely employed to reduce chemotherapy-induced emesis (nausea and vomiting)<sup>251</sup>, to potentiate chemotherapy in lymphoid cancers<sup>246,252</sup> and to counteract inflammatory or autoimmune responses to immunotherapy<sup>253</sup>. Use of synthetic glucocorticoids in solid malignancies, which may or may not express GRs, could jeopardize adjuvant treatments and promote cancer progression. Indeed, in patients with non-small cell lung cancer, synthetic glucocorticoid use predicted decreased response to immune checkpoint inhibitors (including anti-PDL1 immunotherapy), and decreased DFS and overall survival<sup>254–256</sup>.

## Conclusions and perspectives

Although the evidence that stress promotes cancer initiation is inconsistent, there is robust evidence that stress can facilitate cancer progression through modulating most hallmarks of cancer. Molecular and systemic mechanisms mediating these effects have been identified in animal studies, and most have been recognized in patients with cancer. SNS-derived adrenergic stress responses, and adrenergic–inflammatory responses in the context of medical procedures, are key mediators of these deleterious effects of stress. The use of synthetic steroids, and stress-induced glucocorticoid release in some models, were also shown to promote cancer progression, and to reduce efficacy of adjuvant therapies. However, it should be noted that animal studies leverage their ability to synchronize stress exposure with specific phases of cancer growth and metastasis that are critically prone to stress. By contrast, epidemiological studies and most clinical trials assessing stress-reducing psychosocial interventions did not focus on stress-prone phases, some of which cannot be identified and addressed clinically. Thus, it is not a surprise that epidemiological and clinical intervention studies have shown small effect

size or mixed outcomes. Importantly, psychological interventions have the potential to individually address patients' unique sources of stress responses, may exert enduring post-treatment effects without drug adverse effects and are feasible in patients with contraindications to drug therapy. Based on our current understanding of cancer biology, stress and the complex interactions between them along critical time frames in the continuum of cancer, we hypothesize that stress-management interventions can reduce cancer recurrence and mortality, especially in patients undergoing curative oncological surgery. To facilitate such beneficial effects, we suggest that stress-management interventions should be tested during critical periods affecting cancer progression, especially the short perioperative period and adjuvant treatments, and compared with other time periods; should be accompanied by pharmacological approaches to overcome stress and inflammatory responses that are unavoidably triggered by medical procedures; and should include individualized modules to accommodate patient-unique characteristics and needs, and focus on patients with higher manifestation of stress symptomology. Such studies should be powered similarly to testing a new drug therapy, and will likely require prioritization by non-profit funding organizations. Recent biomarker clinical trials, including pharmacological stress-reducing interventions, indicate the potential capacity of such approaches to reduce cancer mortality. Based on the current data, we believe that such approaches should be tested through large collaborative multicentre RCTs, assessing the impact of unified interventions on long-term cancer outcomes, with similar rigour to that employed when studying a new agent for cancer therapy.

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- LeShan, L. Psychological states as factors in the development of malignant disease: a critical review. *J. Natl Cancer Inst.* **22**, 1–18 (1959).
- Mravec, B., Tibensky, M. & Horvathova, L. Stress and cancer. Part I: mechanisms mediating the effect of stressors on cancer. *J. Neuroimmunol.* **346**, 577311 (2020).  
**This review describes mechanisms by which stress affects specific hallmarks of cancer, emphasizing how stress is an integral part of cancer biology.**
- Cole, S. W. & Sood, A. K. Molecular pathways:  $\beta$ -adrenergic signaling in cancer. *Clin. Cancer Res.* **18**, 1201–1206 (2012).
- Eng, J. W.-L. et al. A nervous tumor microenvironment: the impact of adrenergic stress on cancer cells, immunosuppression, and immunotherapeutic response. *Cancer Immunol. Immunother.* **63**, 1115–1128 (2014).
- Armaiz-Pena, G. N., Cole, S. W., Lutgendorf, S. K. & Sood, A. K. Neuroendocrine influences on cancer progression. *Brain Behav. Immun.* **30**, S19–S25 (2013).
- Cole, S. W., Nagaraja, A. S., Lutgendorf, S. K., Green, P. A. & Sood, A. K. Sympathetic nervous system regulation of the tumour microenvironment. *Nat. Rev. Cancer* **15**, 563 (2015).  
**This review describes the contribution of adrenergic signalling to cancer progression, focusing on the tumour microenvironment.**
- Armaiz-Pena, G. N., Colon-Echevarria, C. B. & Lamboy-Caraballo, R. Neuroendocrine regulation of tumor-associated immune cells. *Front. Oncol.* **9**, 1077 (2019).  
**This review examines the effects of sympathetic and/or glucocorticoid signalling on various tumour-associated immune cells.**
- Antoni, M. H. & Dhabhar, F. S. The impact of psychosocial stress and stress management on immune responses in patients with cancer. *Cancer* **125**, 1417–1431 (2019).  
**This review discusses both preclinical and clinical studies and summarizes the effects of stress and stress management on immune indices in cancer, suggesting potential optimal strategies for stress management in patients with cancer.**
- Neeman, E. & Ben-Eliyahu, S. Surgery and stress promote cancer metastasis: new outlooks on perioperative mediating mechanisms and immune involvement. *Brain Behav. Immun.* **30**, S32–S40 (2013).
- Cui, B. et al. Cancer and stress: NextGen strategies. *Brain Behav. Immun.* **93**, 368–383 (2020).
- Lutgendorf, S. K. & Andersen, B. L. Biobehavioral approaches to cancer progression and survival: mechanisms and interventions. *Am. Psychol.* **70**, 186–197 (2015).
- Mravec, B., Tibensky, M. & Horvathova, L. Stress and cancer. Part II: therapeutic implications for oncology. *J. Neuroimmunol.* **346**, 577312 (2020).
- Moreno-Smith, M., Lutgendorf, S. K. & Sood, A. K. Impact of stress on cancer metastasis. *Future Oncol.* **6**, 1863–1881 (2010).
- Selye, H. *The Stress of Life* (McGraw-Hill, 1956).
- Cacioppo, J. T., Cacioppo, S., Capitanio, J. P. & Cole, S. W. The neuroendocrinology of social isolation. *Annu. Rev. Psychol.* **66**, 733–767 (2015).
- Bortolato, B. et al. Depression in cancer: the many biobehavioral pathways driving tumor progression. *Cancer Treat. Rev.* **52**, 58–70 (2017).
- Liu, R. T. & Alloy, L. B. Stress generation in depression: a systematic review of the empirical literature and recommendations for future study. *Clin. Psychol. Rev.* **30**, 582–593 (2010).
- Wang, Q., Timberlake, M. A. II, Prall, K. & Dwivedi, Y. The recent progress in animal models of depression. *Prog. Neuropsychopharmacol. Biol. Psychiatry* **77**, 99–109 (2017).
- Sapolsky, R. M. Stress and the brain: individual variability and the inverted-U. *Nat. Neurosci.* **18**, 1344 (2015).
- McEwen, B. S. Neurobiological and systemic effects of chronic stress. *Chronic Stress* **1**, 2470547017692328 (2017).
- McEwen, B. S. & Stellar, E. Stress and the individual: mechanisms leading to disease. *Arch. Intern. Med.* **153**, 2093–2101 (1993).
- McEwen, B. S. & Gianaros, P. J. Central role of the brain in stress and adaptation: links to socioeconomic status, health, and disease. *Ann. NY Acad. Sci.* **1186**, 190 (2010).
- Lazarus, R. S. & Folkman, S. *Stress, Appraisal, and Coping* (Springer, 1984).
- Holmes, T. H. & Rahe, R. H. The social readjustment rating scale. *J. Psychosom. Res.* **11**, 213–218 (1967).
- McEwen, B. S., Gray, J. D. & Nasca, C. Recognizing resilience: learning from the effects of stress on the brain. *Neurobiol. Stress.* **1**, 1–11 (2015).
- Fava, G. A. et al. Clinical characterization of allostatic overload. *Psychoneuroendocrinology* **108**, 94–101 (2019).
- Kiecolt-Glaser, J. K., Renna, M. E., Shroot, M. R. & Madison, A. A. Stress reactivity: what pushes us higher, faster, and longer — and why it matters. *Curr. Dir. Psychol. Sci.* **29**, 492–498 (2020).
- Hanahan, D. & Weinberg, R. A. Hallmarks of cancer: the next generation. *Cell* **144**, 646–674 (2011).

29. Fouad, Y. A. & Aanei, C. Revisiting the hallmarks of cancer. *Am. J. Cancer Res.* **7**, 1016 (2017).
30. Manjili, M. H. Tumor dormancy and relapse: from a natural byproduct of evolution to a disease state. *Cancer Res.* **77**, 2564–2569 (2017).
31. Bergers, G. & Benjamin, L. E. Tumorigenesis and the angiogenic switch. *Nat. Rev. Cancer* **3**, 401–410 (2003).
32. Patidar, A. et al. DAMP–TLR–cytokine axis dictates the fate of tumor. *Cytokine* **104**, 114–123 (2018).
33. Melamed, R. et al. Marginating pulmonary-NK activity and resistance to experimental tumor metastasis: suppression by surgery and the prophylactic use of a  $\beta$ -adrenergic antagonist and a prostaglandin synthesis inhibitor. *Brain Behav. Immun.* **19**, 114–126 (2005).
34. Melamed, R. et al. The marginating-pulmonary immune compartment in rats: characteristics of continuous inflammation and activated NK cells. *J. Immunother.* **33**, 16–29 (2010).
35. Sorski, L. et al. Prevention of liver metastases through perioperative acute CpG-immune stimulation. *Cancer Immunol. Immunother.* **69**, 2021–2031 (2020).
36. Strlic, B. & Offermanns, S. Intravascular survival and extravasation of tumor cells. *Cancer Cell* **32**, 282–293 (2017).
37. Shaashua, L. et al. Spontaneous regression of micro-metastases following primary tumor excision: a critical role for primary tumor secretome. *BMC Biol.* **18**, 1–13 (2020).
38. Gonzalez, H., Hagerling, C. & Werb, Z. Roles of the immune system in cancer: from tumor initiation to metastatic progression. *Genes Dev.* **32**, 1267–1284 (2018).
39. Rosenne, E. et al. Inducing a mode of NK-resistance to suppression by stress and surgery: a potential approach based on low dose of poly I-C to reduce postoperative cancer metastasis. *Brain Behav. Immun.* **21**, 395–408 (2007).
40. Ben-Eliyahu, S., Shakhari, G., Page, G. G., Stefanski, V. & Shakhari, K. Suppression of NK cell activity and of resistance to metastasis by stress: a role for adrenal catecholamines and  $\beta$ -adrenoceptors. *Neuroimmunomodulation* **8**, 154–164 (2000).
41. Sloan, E. K. et al. The sympathetic nervous system induces a metastatic switch in primary breast cancer. *Cancer Res.* **70**, 7042–7052 (2010).
- This preclinical study in a breast cancer model reports that whereas chronic stress does not promote primary tumour growth, it promotes its metastatic dissemination, demonstrating specific interactions between stress and unique stages in cancer progression.**
42. Du, P. et al. Chronic stress promotes EMT-mediated metastasis through activation of STAT3 signaling pathway by miR-337-3p in breast cancer. *Cell Death Dis.* **11**, 1–13 (2020).
43. Madden, K. S., Szpunar, M. J. & Brown, E. B. Early impact of social isolation and breast tumor progression in mice. *Brain Behav. Immun.* **30**, S135–S141 (2013).
44. Volden, P. A. & Conzen, S. D. The influence of glucocorticoid signaling on tumor progression. *Brain Behav. Immun.* **30**, S26–S31 (2013).
45. Flint, M. S., Baum, A., Chambers, W. H. & Jenkins, F. J. Induction of DNA damage, alteration of DNA repair and transcriptional activation by stress hormones. *Psychoneuroendocrinology* **32**, 470–479 (2007).
46. Hara, M. R. et al. A stress response pathway regulates DNA damage through  $\beta_2$ -adrenoceptors and  $\beta$ -arrestin-1. *Nature* **477**, 349–353 (2011).
47. Hara, M. R., Sachs, B. D., Caron, M. G. & Lefkowitz, R. J. Pharmacological blockade of a  $\beta_2$ AR– $\beta$ -arrestin-1 signaling cascade prevents the accumulation of DNA damage in a behavioral stress model. *Cell Cycle* **12**, 219–224 (2013).
48. Feng, Z. et al. Chronic restraint stress attenuates p53 function and promotes tumorigenesis. *Proc. Natl Acad. Sci. USA* **109**, 7013–7018 (2012).
49. Gidron, Y., Russ, K., Tisarchondou, H. & Warner, J. The relation between psychological factors and DNA-damage: a critical review. *Biol. Psychol.* **72**, 291–304 (2006).
50. Lamboy-Caraballo, R. et al. Norepinephrine-induced DNA damage in ovarian cancer cells. *Int. J. Mol. Sci.* **21**, 2250 (2020).
51. Flaherty, R. L. et al. Glucocorticoids induce production of reactive oxygen species/reactive nitrogen species and DNA damage through an iNOS mediated pathway in breast cancer. *Breast Cancer Res.* **19**, 1–13 (2017).
52. Reeder, A. et al. Stress hormones reduce the efficacy of paclitaxel in triple negative breast cancer through induction of DNA damage. *Br. J. Cancer* **112**, 1461–1470 (2015).
53. Plummer, M. et al. Global burden of cancers attributable to infections in 2012: a synthetic analysis. *Lancet Glob. Health* **4**, e609–e616 (2016).
54. de Martel, C., Georges, D., Bray, F., Ferlay, J. & Clifford, G. M. Global burden of cancer attributable to infections in 2018: a worldwide incidence analysis. *Lancet Glob. Health* **8**, e180–e190 (2020).
55. Antoni, M. H. et al. The influence of bio-behavioural factors on tumour biology: pathways and mechanisms. *Nat. Rev. Cancer* **6**, 240–248 (2006).
56. Irwin, M. R. & Cole, S. W. Reciprocal regulation of the neural and innate immune systems. *Nat. Rev. Immunol.* **11**, 625–632 (2011).
57. Collado-Hidalgo, A., Sung, C. & Cole, S. Adrenergic inhibition of innate anti-viral response: PKA blockade of type I interferon gene transcription mediates catecholamine support for HIV-1 replication. *Brain Behav. Immun.* **20**, 552–563 (2006).
58. Cacioppo, J. T. et al. Autonomic and glucocorticoid associations with the steady-state expression of latent Epstein–Barr virus. *Hormones Behav.* **42**, 32–41 (2002).
59. Glaser, R. et al. The differential impact of training stress and final examination stress on herpesvirus latency at the United States Military Academy at West Point. *Brain Behav. Immun.* **13**, 240–251 (1999).
60. Fang, C. Y. et al. Perceived stress is associated with impaired T-cell response to HPV16 in women with cervical dysplasia. *Ann. Behav. Med.* **35**, 87–96 (2008).
61. Fang, F. et al. Risk of infection-related cancers after the loss of a child: a follow-up study in Sweden. *Cancer Res.* **71**, 116–122 (2011).
62. Saul, A. N. et al. Chronic stress and susceptibility to skin cancer. *J. Natl Cancer Inst.* **97**, 1760–1767 (2005).
63. Sumis, A. et al. Social isolation induces autophagy in the mouse mammary gland: link to increased mammary cancer risk. *Endocr. Relat. Cancer* **23**, 839–856 (2016).
64. Kokolus, K. M. et al. Baseline tumor growth and immune control in laboratory mice are significantly influenced by subthermoneutral housing temperature. *Proc. Natl Acad. Sci. USA* **110**, 20176–20181 (2013).
65. Renz, B. W. et al.  $\beta_2$  adrenergic-neurotrophin feedforward loop promotes pancreatic cancer. *Cancer Cell* **33**, 75–90.e7 (2018).
- This research demonstrates the reciprocal relations between the malignant tissue and its direct sympathetic innervation in preclinical pancreatic cancer models.**
66. Magnon, C. et al. Autonomic nerve development contributes to prostate cancer progression. *Science* **341**, 1236361 (2013).
67. Hermes, G. L. et al. Social isolation dysregulates endocrine and behavioral stress while increasing malignant burden of spontaneous mammary tumors. *Proc. Natl Acad. Sci. USA* **106**, 22393–22398 (2009).
68. Hasen, N. S., O’Leary, K. A., Auger, A. P. & Schuler, L. A. Social isolation reduces mammary development, tumor incidence, and expression of epigenetic regulators in wild-type and p53-heterozygous mice. *Cancer Prev. Res.* **3**, 620–629 (2010).
69. Nguyen, K. D. et al. Alternatively activated macrophages produce catecholamines to sustain adaptive thermogenesis. *Nature* **480**, 104–108 (2011).
70. Flierl, M. A. et al. Phagocyte-derived catecholamines enhance acute inflammatory injury. *Nature* **449**, 721–725 (2007).
71. Wong, H. P. S. et al. Nicotine promotes cell proliferation via  $\alpha 7$ -nicotinic acetylcholine receptor and catecholamine-synthesizing enzymes-mediated pathway in human colon adenocarcinoma HT-29 cells. *Toxicol. Appl. Pharmacol.* **221**, 261–267 (2007).
72. Shi, M. et al. The  $\beta_2$ -adrenergic receptor and Her2 comprise a positive feedback loop in human breast cancer cells. *Breast Cancer Res. Treat.* **125**, 351–362 (2011).
73. Amaro, F. et al.  $\beta$ -Adrenoceptor activation in breast MCF-10A cells induces a pattern of catecholamine production similar to that of tumorigenic MCF-7 cells. *Int. J. Mol. Sci.* **21**, 7968 (2020).
74. Zhang, X. et al. Chronic stress promotes gastric cancer progression and metastasis: an essential role for ADRB2. *Cell Death Dis.* **10**, 1–15 (2019).
75. Zhi, X. et al. Adrenergic modulation of AMPK-dependent autophagy by chronic stress enhances cell proliferation and survival in gastric cancer. *Int. J. Oncol.* **54**, 1625–1638 (2019).
76. Wong, H. P. et al. Effects of adrenaline in human colon adenocarcinoma HT-29 cells. *Life Sci.* **88**, 1108–1112 (2011).
77. Sood, A. K. et al. Adrenergic modulation of focal adhesion kinase protects human ovarian cancer cells from anoikis. *J. Clin. Invest.* **120**, 1515–1523 (2010).
78. Liu, H. et al. Activation of adrenergic receptor  $\beta_2$  promotes tumor progression and epithelial mesenchymal transition in tongue squamous cell carcinoma. *Int. J. Mol. Med.* **41**, 147–154 (2018).
79. Nagaraja, A. S. et al. Sustained adrenergic signaling leads to increased metastasis in ovarian cancer via increased PGE<sub>2</sub> synthesis. *Oncogene* **35**, 2390–2397 (2016).
80. Kim-Fuchs, C. et al. Chronic stress accelerates pancreatic cancer growth and invasion: a critical role for  $\beta$ -adrenergic signaling in the pancreatic microenvironment. *Brain Behav. Immun.* **40**, 40–47 (2014).
81. Moretti, S. et al.  $\beta$ -Adrenoceptors are upregulated in human melanoma and their activation releases pro-tumorigenic cytokines and metalloproteases in melanoma cell lines. *Lab. Invest.* **93**, 279–290 (2013).
82. Pu, J. et al. Adrenaline promotes epithelial-to-mesenchymal transition via HuR–TGF $\beta$  regulatory axis in pancreatic cancer cells and the implication in cancer prognosis. *Biochem. Biophys. Res. Commun.* **493**, 1273–1279 (2017).
83. Liu, J. et al. A novel  $\beta_2$ -AR/YB-1/ $\beta$ -catenin axis mediates chronic stress-associated metastasis in hepatocellular carcinoma. *Oncogenesis* **9**, 1–14 (2020).
84. Bucsek, M. J. et al.  $\beta$ -Adrenergic signaling in mice housed at standard temperatures suppresses an effector phenotype in CD8<sup>+</sup> T cells and undermines checkpoint inhibitor therapy. *Cancer Res.* **77**, 5639–5651 (2017).
85. Chen, H. et al. Chronic psychological stress promotes lung metastatic colonization of circulating niche cancer cells by decorating a pre-metastatic nest through activating  $\beta$ -adrenergic signaling. *J. Pathol.* **244**, 49–60 (2018).
86. Lamkin, D. M. et al. Chronic stress enhances progression of acute lymphoblastic leukemia via  $\beta$ -adrenergic signaling. *Brain Behav. Immun.* **26**, 635–641 (2012).
87. Thaker, P. H. et al. Chronic stress promotes tumor growth and angiogenesis in a mouse model of ovarian carcinoma. *Nat. Med.* **12**, 939–944 (2006).
- This is the first preclinical study to demonstrate the effects of chronic stress on tumour angiogenesis, which also identifies the mediating adrenergic signalling pathway.**
88. Chang, A. et al.  $\beta_2$ -Adrenoceptors on tumor cells play a critical role in stress-enhanced metastasis in a mouse model of breast cancer. *Brain Behav. Immun.* **57**, 106–115 (2016).
89. Zahalka, A. H. & Frenette, P. S. Nerves in cancer. *Nat. Rev. Cancer* **20**, 143–157 (2020).
90. Qin, J.-F. et al. Adrenergic receptor  $\beta_2$  activation by stress promotes breast cancer progression through macrophages M2 polarization in tumor microenvironment. *BMB Rep.* **48**, 295 (2015).
91. Lutgendorf, S. K. et al. Social isolation is associated with elevated tumor norepinephrine in ovarian carcinoma patients. *Brain Behav. Immun.* **25**, 250–255 (2011).
92. Yang, H. et al. Stress–glucocorticoid–TSC22D3 axis compromises therapy-induced antitumor immunity. *Nat. Med.* **25**, 1428–1441 (2019).
- This preclinical study conducted in several cancer models identifies a novel stress-induced mechanism, mediated through glucocorticoid signalling in dendritic cells, that can compromise chemotherapy-induced and immunotherapy-induced antitumour immunity.**
93. Obradović, M. M. et al. Glucocorticoids promote breast cancer metastasis. *Nature* **567**, 540–544 (2019).
- This preclinical study uses several models of breast cancer to demonstrate that the activation of GR in breast cancer cells, through ROR1 kinase signalling, leads to increased metastasis and resistance to chemotherapy, thus emphasizing that GR signalling, either by endogenous (stress-induced) or exogenous sources of glucocorticoids, can worsen cancer progression.**
94. Pan, D., Kocherginsky, M. & Conzen, S. D. Activation of the glucocorticoid receptor is associated with poor



- prognosis in estrogen receptor-negative breast cancer. *Cancer Res.* **71**, 6360–6370 (2011).
95. Madden, K. S., Szpunar, M. J. & Brown, E. B.  $\beta$ -Adrenergic receptors ( $\beta$ -AR) regulate VEGF and IL-6 production by divergent pathways in high  $\beta$ -AR-expressing breast cancer cell lines. *Breast Cancer Res. Treat.* **130**, 747–758 (2011).
96. Lutgendorf, S. K. et al. Stress-related mediators stimulate vascular endothelial growth factor secretion by two ovarian cancer cell lines. *Clin. Cancer Res.* **9**, 4514–4521 (2003).
97. Yang, E. V. et al. Norepinephrine upregulates VEGF, IL-8, and IL-6 expression in human melanoma tumor cell lines: implications for stress-related enhancement of tumor progression. *Brain Behav. Immun.* **23**, 267–275 (2009).
98. Chen, H. et al. Adrenergic signaling promotes angiogenesis through endothelial cell–tumor cell crosstalk. *Endocr. Relat. Cancer* **21**, 783–795 (2014).
99. Shan, T. et al.  $\beta$ 2-AR–HIF-1 $\alpha$ : a novel regulatory axis for stress-induced pancreatic tumor growth and angiogenesis. *Curr. Mol. Med.* **13**, 1023–1034 (2013).
100. Xu, P. et al. Surgical trauma contributes to progression of colon cancer by downregulating CXCL4 and recruiting MDSCs. *Exp. Cell Res.* **370**, 692–698 (2018).
101. Budiu, R. A. et al. Restraint and social isolation stressors differentially regulate adaptive immunity and tumor angiogenesis in a breast cancer mouse model. *Cancer Clin. Oncol.* **6**, 12 (2017).
102. Hulsurkar, M. et al.  $\beta$ -Adrenergic signaling promotes tumor angiogenesis and prostate cancer progression through HDAC2-mediated suppression of thrombospondin-1. *Oncogene* **36**, 1525–1536 (2017).
103. Lutgendorf, S. K. et al. Vascular endothelial growth factor and social support in patients with ovarian carcinoma. *Cancer* **95**, 808–815 (2002).
104. Lutgendorf, S. K. et al. Biobehavioral influences on matrix metalloproteinase expression in ovarian carcinoma. *Clin. Cancer Res.* **14**, 6839–6846 (2008).
105. Costanzo, E. S. et al. Psychosocial factors and interleukin-6 among women with advanced ovarian cancer. *Cancer* **104**, 305–313 (2005).
106. Lutgendorf, S. K. et al. Interleukin-6, cortisol, and depressive symptoms in ovarian cancer patients. *J. Clin. Oncol.* **26**, 4820–4827 (2008).
107. Stack, S. A. et al. Lymphangiogenesis and lymphatic vessel remodelling in cancer. *Nat. Rev. Cancer* **14**, 159–172 (2014).
108. Le, C. P. et al. Chronic stress in mice remodels lymph vasculature to promote tumour cell dissemination. *Nat. Commun.* **7**, 10634 (2016).
- This preclinical study demonstrates the effects of chronic stress on lymphatic modulation and metastasis, and identifies the underlying adrenergic mechanisms.**
109. Bower, J. E. et al. Prometastatic molecular profiles in breast tumors from socially isolated women. *JNCI Cancer Spectr.* **2**, pky029 (2018).
110. Qiao, G., Chen, M., Bucsek, M. J., Repasky, E. A. & Hylander, B. L. Adrenergic signaling: a targetable checkpoint limiting development of the antitumor immune response. *Front. Immunol.* **9**, 164 (2018).
111. Hirata, T. & Narumiya, S. (2012). In *Advances in Immunology* (ed. Alt, F. W.) 143–174 (Elsevier, 2012).
112. Shakhbar, G. & Ben-Eliyahu, S. In vivo  $\beta$ -adrenergic stimulation suppresses natural killer activity and compromises resistance to tumor metastasis in rats. *J. Immunol.* **160**, 3251–3258 (1998).
113. Inbar, S. et al. Do stress responses promote leukemia progression? An animal study suggesting a role for epinephrine and prostaglandin-E<sub>2</sub> through reduced NK activity. *PLoS ONE* **6**, e19246 (2011).
114. Rosenne, E. et al. In vivo suppression of NK cell cytotoxicity by stress and surgery: glucocorticoids have a minor role compared to catecholamines and prostaglandins. *Brain Behav. Immun.* **37**, 207–219 (2014).
115. Lutgendorf, S. K. et al. Social support, psychological distress, and natural killer cell activity in ovarian cancer. *J. Clin. Oncol.* **23**, 7105–7113 (2005).
116. Hou, N. et al. A novel chronic stress-induced shift in the T<sub>H</sub>1 to T<sub>H</sub>2 response promotes colon cancer growth. *Biochem. Biophys. Res. Commun.* **439**, 471–476 (2013).
117. Lutgendorf, S. K. et al. Depressed and anxious mood and T-cell cytokine expressing populations in ovarian cancer patients. *Brain Behav. Immun.* **22**, 890–900 (2008).
118. Mohamadpour, H. et al.  $\beta$ 2 adrenergic receptor-mediated signaling regulates the immunosuppressive potential of myeloid-derived suppressor cells. *J. Clin. Invest.* **129**, 5537–5552 (2019).
119. Mundy-Bosse, B. L., Thornton, L. M., Yang, H.-C., Andersen, B. L. & Carson, W. E. Psychological stress is associated with altered levels of myeloid-derived suppressor cells in breast cancer patients. *Cell. Immunol.* **270**, 80–87 (2011).
120. Armaiz-Pena, G. N. et al. Adrenergic regulation of monocyte chemotactic protein 1 leads to enhanced macrophage recruitment and ovarian carcinoma growth. *Oncotarget* **6**, 4266–4273 (2015).
121. Lamkin, D. M. et al.  $\beta$ -Adrenergic-stimulated macrophages: comprehensive localization in the M1–M2 spectrum. *Brain Behav. Immun.* **57**, 338–346 (2016).
122. Campbell, J. P. et al. Stimulation of host bone marrow stromal cells by sympathetic nerves promotes breast cancer bone metastasis in mice. *PLoS Biol.* **10**, e1001363 (2012).
123. Simpson, C. D., Anyiwe, K. & Schimmer, A. D. Anoikis resistance and tumor metastasis. *Cancer Lett.* **272**, 177–185 (2008).
124. Lutgendorf, S. K. et al. Epithelial–mesenchymal transition polarization in ovarian carcinomas from patients with high social isolation. *Cancer* **126**, 4407–4413 (2020).
125. Benish, M. et al. Perioperative use of  $\beta$ -blockers and COX-2 inhibitors may improve immune competence and reduce the risk of tumor metastasis. *Ann. Surg. Oncol.* **15**, 2042–2052 (2008).
126. Kaira, K. et al. Prognostic impact of  $\beta$ 2 adrenergic receptor expression in surgically resected pulmonary pleomorphic carcinoma. *Anticancer Res.* **39**, 395–403 (2019).
127. Choy, C. et al. Inhibition of  $\beta$ 2-adrenergic receptor reduces triple-negative breast cancer brain metastases: the potential benefit of perioperative  $\beta$ -blockade. *Oncol. Rep.* **35**, 3135–3142 (2016).
128. Al-Niaimi, A. et al. The impact of perioperative  $\beta$  blocker use on patient outcomes after primary cytoreductive surgery in high-grade epithelial ovarian carcinoma. *Gynecol. Oncol.* **143**, 521–525 (2016).
129. Barron, T. I., Connolly, R. M., Sharp, L., Bennett, K. & Visvanathan, K.  $\beta$  blockers and breast cancer mortality: a population-based study. *J. Clin. Oncol.* **29**, 2635–2644 (2011).
130. Lemeshev, S. et al.  $\beta$ -Blockers and survival among Danish patients with malignant melanoma: a population-based cohort study. *Cancer Epidemiol. Biomarkers Prev.* **20**, 2273–2279 (2011).
131. Cata, J. P. et al. Perioperative  $\beta$ -blocker use and survival in lung cancer patients. *J. Clin. Anesth.* **26**, 106–117 (2014).
132. Heitz, F. et al. Intake of selective  $\beta$  blockers has no impact on survival in patients with epithelial ovarian cancer. *Gynecol. Oncol.* **144**, 181–186 (2017).
133. Matzner, P. et al. Deleterious synergistic effects of distress and surgery on cancer metastasis: abolishment through an integrated perioperative immune-stimulating stress-inflammatory-reducing intervention. *Brain Behav. Immun.* **80**, 170–178 (2019).
134. Stefanski, V. & Ben-Eliyahu, S. Social confrontation and tumor metastasis in rats: defeat and  $\beta$ -adrenergic mechanisms. *Physiol. Behav.* **60**, 277–282 (1996).
135. Dhabhar, F. S. et al. Short-term stress enhances cellular immunity and increases early resistance to squamous cell carcinoma. *Brain Behav. Immun.* **24**, 127–137 (2010).
136. Benaroya-Milshtien, N., Hollander, N., Apter, A., Yaniv, I. & Pick, C. G. Stress conditioning in mice: alterations in immunity and tumor growth. *Stress* **14**, 301–311 (2011).
137. Williams, J. B. et al. A model of gene–environment interaction reveals altered mammary gland gene expression and increased tumor growth following social isolation. *Cancer Prev. Res.* **2**, 850–861 (2009).
138. Dawes, R. P. et al. Chronic stress exposure suppresses mammary tumor growth and reduces circulating exosome TGF- $\beta$  content via  $\beta$ -adrenergic receptor signaling in MMTV-PyMT mice. *Breast Cancer* **14**, 1178223420931511 (2020).
139. Huo, J. et al. Bone marrow-derived mesenchymal stem cells promoted cutaneous wound healing by regulating keratinocyte migration via  $\beta$ 2-adrenergic receptor signaling. *Mol. Pharmaceutics* **15**, 2513–2527 (2018).
140. Ren, H. et al. Inhibition of  $\alpha$ 1-adrenoceptor reduces TGF- $\beta$ 1-induced epithelial-to-mesenchymal transition and attenuates UUO-induced renal fibrosis in mice. *FASEB J.* **34**, 14892–14904 (2020).
141. Panina-Bordignon, P. et al.  $\beta$ 2-agonists prevent T<sub>H</sub>1 development by selective inhibition of interleukin 12. *J. Clin. Invest.* **100**, 1513–1519 (1997).
142. Ağaç, D., Estrada, L. D., Maples, R., Hooper, L. V. & Farrar, J. D. The  $\beta$ 2-adrenergic receptor controls inflammation by driving rapid IL-10 secretion. *Brain Behav. Immun.* **74**, 176–185 (2018).
143. Kavelaars, A., Van De Pol, M., Zijlstra, J. & Heijnen, C. J.  $\beta$ 2-Adrenergic activation enhances interleukin-8 production by human monocytes. *J. Neuroimmunol.* **77**, 211–216 (1997).
144. Steinle, J. J., Cappoccia, F. C. Jr & Jiang, Y.  $\beta$ -Adrenergic receptor regulation of growth factor protein levels in human choroidal endothelial cells. *Growth Factors* **26**, 325–330 (2008).
145. Asano, A., Morimatsu, M., Nikami, H., Yoshida, T. & Saito, M. Adrenergic activation of vascular endothelial growth factor mRNA expression in rat brown adipose tissue: implication in cold-induced angiogenesis. *Biochem. J.* **328**, 179–183 (1997).
146. Chida, Y., Hamer, M., Wardle, J. & Steptoe, A. Do stress-related psychosocial factors contribute to cancer incidence and survival? *Nat. Clin. Pract. Oncol.* **5**, 466–475 (2008).
- This paper is the most comprehensive meta-analysis assessing the contribution of psychosocial stress to cancer incidence, survival and mortality in several human malignancies.**
147. Coyne, J. C., Ranchor, A. V. & Palmer, S. C. Meta-analysis of stress-related factors in cancer. *Nat. Rev. Clin. Oncol.* **7**, 1–2 (2010).
148. Mravec, B. & Tibensky, M. Increased cancer incidence in “cold” countries: an (un)sympathetic connection? *J. Therm. Biol.* **89**, 102538 (2020).
149. Keinan-Boker, L., Vin-Raviv, N., Lipshitz, I., Linn, S. & Barchana, M. Cancer incidence in Israeli Jewish survivors of World War II. *J. Natl. Cancer Inst.* **101**, 1489–1500 (2009).
150. Huang, T. et al. Depression and risk of epithelial ovarian cancer: results from two large prospective cohort studies. *Gynecol. Oncol.* **139**, 481–486 (2015).
151. Schoemaker, M. J. et al. Psychological stress, adverse life events and breast cancer incidence: a cohort investigation in 106,000 women in the United Kingdom. *Breast Cancer Res.* **18**, 72 (2016).
152. Trudel-Fitzgerald, C. et al. The association of work characteristics with ovarian cancer risk and mortality. *Psychosom. Med.* **79**, 1059 (2017).
153. Liang, J.-A. et al. The analysis of depression and subsequent cancer risk in Taiwan. *Cancer Epidemiol. Prev. Biomarkers* **20**, 473–475 (2011).
154. Heikilä, K. et al. Work stress and risk of cancer: meta-analysis of 5700 incident cancer events in 116 000 European men and women. *BMJ* **346**, f165 (2013).
155. Yang, T. et al. Work stress and the risk of cancer: a meta-analysis of observational studies. *Int. J. Cancer* **144**, 2390–2400 (2019).
156. Perego, M. et al. Reactivation of dormant tumor cells by modified lipids derived from stress-activated neutrophils. *Sci. Transl. Med.* **12**, eabb5817 (2020).
- This preclinical study identifies a distinct mechanism by which tumour-associated neutrophils respond to stress-induced adrenergic activation, and lead to reactivation of dormant tumour cells. This study highlights  $\beta$ -blockade as a potential strategy to prevent stress-induced cancer relapse.**
157. Krall, J. A. et al. The systemic response to surgery triggers the outgrowth of distant immune-controlled tumors in mouse models of dormancy. *Sci. Transl. Med.* **10**, ean3464 (2018).
158. Decker, A. M. et al. Sympathetic signaling reactivates quiescent disseminated prostate cancer cells in the bone marrow. *Mol. Cancer Res.* **15**, 1644–1655 (2017).
159. Gil, F., Costa, G., Hilker, I. & Benito, L. First anxiety, afterwards depression: psychological distress in cancer patients at diagnosis and after medical treatment. *Stress. Health* **28**, 362–367 (2012).
160. Carlson, L. et al. High levels of untreated distress and fatigue in cancer patients. *Br. J. Cancer* **90**, 2297–2304 (2004).
161. Wang, X. et al. Prognostic value of depression and anxiety on breast cancer recurrence and mortality: a systematic review and meta-analysis of 282,203 patients. *Mol. Psychiatry* **25**, 3186–3197 (2020).
162. Pinquart, M. & Duberstein, P. Depression and cancer mortality: a meta-analysis. *Psychol. Med.* **40**, 1797 (2010).



163. Pinquart, M. & Duberstein, P. R. Associations of social networks with cancer mortality: a meta-analysis. *Crit. Rev. Oncol. Hematol.* **75**, 122–137 (2010).
164. Cohen, L. et al. Depressive symptoms and cortisol rhythmicity predict survival in patients with renal cell carcinoma: role of inflammatory signaling. *PLoS ONE* **7**, e42324 (2012).
165. Lutgendorf, S. K. et al. Social influences on clinical outcomes of patients with ovarian cancer. *J. Clin. Oncol.* **30**, 2885 (2012).
166. Chou, A. F., Stewart, S. L., Wild, R. C. & Bloom, J. R. Social support and survival in young women with breast carcinoma. *Psychooncology* **21**, 125–133 (2012).
167. Kroenke, C. H. et al. Prediagnosis social support, social integration, living status, and colorectal cancer mortality in postmenopausal women from the women's health initiative. *Cancer* **126**, 1766–1775 (2020).
168. Fagundes, C. P. et al. Basal cell carcinoma: stressful life events and the tumor environment. *Arch. Gen. Psychiatry* **69**, 618–626 (2012).
169. Armer, J. S. et al. Life stress as a risk factor for sustained anxiety and cortisol dysregulation during the first year of survivorship in ovarian cancer. *Cancer* **124**, 3401–3408 (2018).
170. Mirosevic, S. et al. "Not just another meta-analysis": sources of heterogeneity in psychosocial treatment effect on cancer survival. *Cancer Med.* **8**, 363–373 (2019).  
**This meta-analysis assesses the effects of psychosocial stress management on cancer survival, discusses limitations of meta-analytic methods and identifies subpopulations that may better benefit from stress-management approaches.**
171. Fu, W. W. et al. The impact of psychosocial intervention on survival in cancer: a meta-analysis. *Ann. Palliat. Med.* **5**, 93–106 (2016).
172. Xia, Y. et al. Psychosocial and behavioral interventions and cancer patient survival again: hints of an adjusted meta-analysis. *Integr. Cancer Therapies* **13**, 301–309 (2014).
173. Oh, P., Shin, S., Ahn, H. S. & Kim, H. Meta-analysis of psychosocial interventions on survival time in patients with cancer. *Psychol. Health* **31**, 396–419 (2016).
174. Andersen, B. L. et al. Psychological, behavioral, and immune changes after a psychological intervention: a clinical trial. *J. Clin. Oncol.* **22**, 3570 (2004).
175. Fawzy, F. I. et al. A structured psychiatric intervention for cancer patients. I. Changes over time in methods of coping and affective disturbance. *Arch. Gen. Psychiatry* **47**, 720–725 (1990).
176. Antoni, M. H. et al. Cognitive-behavioral stress management reverses anxiety-related leukocyte transcriptional dynamics. *Biol. Psychiatry* **71**, 366–372 (2012).
177. Fawzy, F. I. & Fawzy, N. W. Malignant melanoma: effects of a brief, structured psychiatric intervention on survival and recurrence at 10-year follow-up. *Arch. Gen. Psychiatry* **60**, 100–103 (2003).
178. Stefanek, M. E., Palmer, S. C., Thombs, B. D. & Coyne, J. C. Finding what is not there: unwarranted claims of an effect of psychosocial intervention on recurrence and survival. *Cancer* **115**, 5612–5616 (2009).
179. Coyne, J. C. & Tennen, H. Positive psychology in cancer care: bad science, exaggerated claims, and unproven medicine. *Ann. Behav. Med.* **39**, 16–26 (2010).
180. Coyne, J. C., Stefanek, M. & Palmer, S. C. Psychotherapy and survival in cancer: the conflict between hope and evidence. *Psychol. Bull.* **133**, 367 (2007).
181. Kraemer, H. C., Kuchler, T. & Spiegel, D. Use and misuse of the consolidated standards of reporting trials (CONSORT) guidelines to assess research findings: comment on Coyne, Stefanek, and Palmer (2007). *Psychol. Bull.* **135**, 173–178 (2009).
182. Spiegel, D., Bloom, J. R., Kraemer, H. C. & Gottheil, E. Effect of psychosocial treatment on survival of patients with metastatic breast cancer. *Lancet* **2**, 888–891 (1989).
183. Spiegel, D. et al. Effects of supportive-expressive group therapy on survival of patients with metastatic breast cancer: a randomized prospective trial. *Cancer* **110**, 1130–1138 (2007).
184. Goodwin, P. J. et al. The effect of group psychosocial support on survival in metastatic breast cancer. *N. Engl. J. Med.* **345**, 1719–1726 (2001).
185. Boesen, E. H. et al. Survival after a psychoeducational intervention for patients with cutaneous malignant melanoma: a replication study. *J. Clin. Oncol.* **25**, 5698–5703 (2007).
186. Ben-Eliyahu, S. Tumor excision as a metastatic russian roulette: perioperative interventions to improve long-term survival of cancer patients. *Trends Cancer* **6**, 951–959 (2020).
187. Burton, M. V. et al. A randomized controlled trial of preoperative psychological preparation for mastectomy. *Psychooncology* **4**, 1–19 (1995).
188. Kuchler, T., Bestmann, B., Rappat, S., Henne-Bruns, D. & Wood-Dauphinee, S. Impact of psychotherapeutic support for patients with gastrointestinal cancer undergoing surgery: 10-year survival results of a randomized trial. *J. Clin. Oncol.* **25**, 2702–2708 (2007).
189. Zhang, X.-D. et al. Perioperative comprehensive supportive care interventions for Chinese patients with esophageal carcinoma: a prospective study. *Asian Pac. J. Cancer Prev.* **14**, 7359–7366 (2013).
190. Shaashua, L. et al. Perioperative COX-2 and  $\beta$ -adrenergic blockade improves metastatic biomarkers in breast cancer patients in a phase-II randomized trial. *Clin. Cancer Res.* **23**, 4651–4661 (2017).  
**This study is the first clinical trial in patients with cancer to assess perioperative safety and efficacy of the combined use of propranolol and etodolac on biomarkers related to breast cancer progression.**
191. Benjamin, B. et al. Effect of  $\beta$  blocker combined with COX-2 inhibitor on colonic anastomosis in rats. *Int. J. Colorectal Dis.* **25**, 1459–1464 (2010).
192. Hazut, O. et al. The effect of  $\beta$ -adrenergic blockade and COX-2 inhibition on healing of colon, muscle, and skin in rats undergoing colonic anastomosis. *Int. J. Clin. Pharmacol. Ther.* **49**, 545–554 (2011).
193. Hiller, J. G. et al. Preoperative  $\beta$ -blockade with propranolol reduces biomarkers of metastasis in breast cancer: a phase II randomized trial. *Clin. Cancer Res.* **26**, 1803–1811 (2020).
194. Jang, H. I., Lim, S. H., Lee, Y. Y., Kim, T. J. & Choi, C. H. Perioperative administration of propranolol to women undergoing ovarian cancer surgery: a pilot study. *Obstet. Gynecol. Sci.* **60**, 170–177 (2017).
195. Knight, J. M. et al. Propranolol inhibits molecular risk markers in HCT recipients: a phase 2 randomized controlled biomarker trial. *Blood Adv.* **4**, 467–476 (2020).
196. Gandhi, S. et al. Phase I clinical trial of combination propranolol and pembrolizumab in locally advanced and metastatic melanoma: safety, tolerability, and preliminary evidence of antitumor activity. *Clin. Cancer Res.* **27**, 87–95 (2021).
197. Ricon, I., Hanalis-Miller, T., Haldar, R., Jacoby, R. & Ben-Eliyahu, S. Perioperative biobehavioral interventions to prevent cancer recurrence through combined inhibition of  $\beta$ -adrenergic and cyclooxygenase 2 signaling. *Cancer* **125**, 45–56 (2019).
198. Horowitz, M., Neeman, E., Sharon, E. & Ben-Eliyahu, S. Exploiting the critical perioperative period to improve long-term cancer outcomes. *Nat. Rev. Clin. Oncol.* **12**, 213–226 (2015).  
**This review summarizes important aspects within the perioperative period that make this time frame critical in affecting long-term cancer outcomes, and suggests potential clinical perioperative interventions to reduce metastatic disease.**
199. Hiller, J. G., Perry, N. J., Poulgiannis, G., Riedel, B. & Sloan, E. K. Perioperative events influence cancer recurrence risk after surgery. *Nat. Rev. Clin. Oncol.* **15**, 205–218 (2018).  
**This review highlights perioperative events as critical in affecting cancer outcomes and suggests how to reduce perioperative risks.**
200. Sorski, L. et al. Reducing liver metastases of colon cancer in the context of extensive and minor surgeries through  $\beta$ -adrenoceptors blockade and COX2 inhibition. *Brain Behav. Immun.* **58**, 91–98 (2016).
201. Glasner, A. et al. Improving survival rates in two models of spontaneous postoperative metastasis in mice by combined administration of a  $\beta$ -adrenergic antagonist and a cyclooxygenase-2 inhibitor. *J. Immunol.* **184**, 2449–2457 (2010).  
**This preclinical study demonstrates the synergistic beneficial effects of perioperative blockade of adrenergic and prostaglandin signalling on immunity and postoperative survival in two models of spontaneous metastasis.**
202. Haldar, R. et al. Perioperative inhibition of  $\beta$ -adrenergic and COX2 signaling in a clinical trial in breast cancer patients improves tumor Ki-67 expression, serum cytokine levels, and PBMCs transcriptome. *Brain Behav. Immun.* **73**, 294–309 (2018).
203. Haldar, R. et al. Perioperative COX2 and  $\beta$ -adrenergic blockade improves biomarkers of tumor metastasis, immunity, and inflammation in colorectal cancer: a randomized controlled trial. *Cancer* **126**, 3991–4001 (2020).  
**This clinical trial demonstrates safety, feasibility and efficacy of perioperative combined treatment with propranolol and etodolac to improve cancer biomarkers and, potentially, survival outcomes in patients with CRC.**
204. US National Library of Medicine. *ClinicalTrials.gov* <https://clinicaltrials.gov/ct2/show/NCT03838029> (2019).
205. US National Library of Medicine. *ClinicalTrials.gov* <https://clinicaltrials.gov/ct2/show/NCT03919461> (2019).
206. Busby, J., Mills, K., Zhang, S.-D., Liberante, F. G. & Cardwell, C. R. Selective serotonin reuptake inhibitor use and breast cancer survival: a population-based cohort study. *Breast Cancer Res.* **20**, 4 (2018).
207. Boursi, B., Lurie, I., Haynes, K., Mamtani, R. & Yang, Y.-X. Chronic therapy with selective serotonin reuptake inhibitors and survival in newly diagnosed cancer patients. *Eur. J. Cancer Care* **27**, e12666 (2018).
208. Zingone, A. et al. Relationship between anti-depressant use and lung cancer survival. *Cancer Treat. Res. Commun.* **10**, 33–39 (2017).
209. Stockler, M. R. et al. Effect of sertraline on symptoms and survival in patients with advanced cancer, but without major depression: a placebo-controlled double-blind randomised trial. *Lancet Oncol.* **8**, 603–612 (2007).
210. Sternbach, H. Are antidepressants carcinogenic? A review of preclinical and clinical studies. *J. Clin. Psychiatry* **64**, 1153–1162 (2003).
211. Grygier, B. et al. Inhibitory effect of antidepressants on B16F10 melanoma tumor growth. *Pharmacol. Rep.* **65**, 672–681 (2013).
212. Kubera, M. et al. Stimulatory effect of antidepressant drug pretreatment on progression of B16F10 melanoma in high-active male and female C57BL/6J mice. *J. Neuroimmunol.* **240–241**, 34–44 (2011).
213. Andersen, B. L., Shapiro, C. L., Farrar, W. B., Crespin, T. & Wells-DiGregorio, S. Psychological responses to cancer recurrence: a controlled prospective study. *Cancer* **104**, 1540–1547 (2005).
214. Linden, W., Vodermaier, A., MacKenzie, R. & Greig, D. Anxiety and depression after cancer diagnosis: prevalence rates by cancer type, gender, and age. *J. Affect. Disord.* **141**, 343–351 (2012).
215. Mitchell, A. J., Ferguson, D. W., Gill, J., Paul, J. & Symonds, P. Depression and anxiety in long-term cancer survivors compared with spouses and healthy controls: a systematic review and meta-analysis. *Lancet Oncol.* **14**, 721–732 (2013).
216. Watts, S., Prescott, P., Mason, J., McLeod, N. & Lewith, G. Depression and anxiety in ovarian cancer: a systematic review and meta-analysis of prevalence rates. *BMJ Open* **5**, e007618 (2015).
217. Sephton, S. E. et al. Depression, cortisol, and suppressed cell-mediated immunity in metastatic breast cancer. *Brain Behav. Immun.* **23**, 1148–1155 (2009).
218. Andersen, B. L. et al. Stress and immune responses after surgical treatment for regional breast cancer. *J. Natl Cancer Inst.* **90**, 30–36 (1998).
219. Blomberg, B. B. et al. Psychosocial adaptation and cellular immunity in breast cancer patients in the weeks after surgery: an exploratory study. *J. Psychosom. Res.* **67**, 369–376 (2009).
220. Schrepf, A. et al. Cortisol and inflammatory processes in ovarian cancer patients following primary treatment: relationships with depression, fatigue, and disability. *Brain Behav. Immun.* **30**, S126–S134 (2013).
221. Pyter, L. M., Pinerov, V., Galang, J. A., McClintock, M. K. & Prendergast, B. J. Peripheral tumors induce depressive-like behaviors and cytokine production and alter hypothalamic–pituitary–adrenal axis regulation. *Proc. Natl Acad. Sci. USA* **106**, 9069–9074 (2009).
222. Bower, J. E. & Lamkin, D. M. Inflammation and cancer-related fatigue: mechanisms, contributing factors, and treatment implications. *Brain Behav. Immun.* **30**, S48–S57 (2013).
223. Bower, J. E. et al. Inflammation and behavioral symptoms after breast cancer treatment: do fatigue, depression, and sleep disturbance share a common underlying mechanism? *J. Clin. Oncol.* **29**, 3517 (2011).
224. Norden, D. M. et al. Tumor growth increases neuroinflammation, fatigue and depressive-like

- behavior prior to alterations in muscle function. *Brain Behav. Immun.* **43**, 76–85 (2015).
225. Vardy, J. L. et al. Cognitive function in patients with colorectal cancer who do and do not receive chemotherapy: a prospective, longitudinal, controlled study. *J. Clin. Oncol.* **33**, 4085 (2015).
226. Hutchinson, A. D., Hosking, J. R., Kichenadasse, G., Mattiske, J. K. & Wilson, C. Objective and subjective cognitive impairment following chemotherapy for cancer: a systematic review. *Cancer Treat. Rev.* **38**, 926–934 (2012).
227. Chrousos, G. P. The hypothalamic–pituitary–adrenal axis and immune-mediated inflammation. *N. Engl. J. Med.* **332**, 1351–1363 (1995).
228. Ben-Shaanan, T. L. et al. Modulation of anti-tumor immunity by the brain's reward system. *Nat. Commun.* **9**, 2723 (2018).
229. Matzner, P. et al. Harnessing cancer immunotherapy during the unexploited immediate perioperative period. *Nat. Rev. Clin. Oncol.* **17**, 313–326 (2020).
230. Goldfarb, Y. et al. Improving postoperative immune status and resistance to cancer metastasis: a combined perioperative approach of immunostimulation and prevention of excessive surgical stress responses. *Ann. Surg.* **253**, 798–810 (2011).
231. Levi, B. et al. Stress impairs the efficacy of immune stimulation by CpG-C: potential neuroendocrine mediating mechanisms and significance to tumor metastasis and the perioperative period. *Brain Behav. Immun.* **56**, 209–220 (2016).
232. Shaashua, L. et al. Plasma IL-12 levels are suppressed in vivo by stress and surgery through endogenous release of glucocorticoids and prostaglandins but not catecholamines or opioids. *Psychoneuroendocrinology* **42**, 11–23 (2014).
233. Sommershof, A., Scheuermann, L., Koerner, J. & Groettrup, M. Chronic stress suppresses anti-tumor T CD8<sup>+</sup> responses and tumor regression following cancer immunotherapy in a mouse model of melanoma. *Brain Behav. Immun.* **65**, 140–149 (2017).
234. Nissen, M. D., Sloan, E. K. & Mattarollo, S. R.  $\beta$ -adrenergic signaling impairs antitumor CD8<sup>+</sup> T-cell responses to B-cell lymphoma immunotherapy. *Cancer Immunol. Res.* **6**, 98–109 (2018).
235. Kang, Y. et al. Adrenergic stimulation of DUSP1 impairs chemotherapy response in ovarian cancer. *Clin. Cancer Res.* **22**, 1713–1724 (2016).
236. Deng, G.-H. et al. Exogenous norepinephrine attenuates the efficacy of sunitinib in a mouse cancer model. *J. Exp. Clin. Cancer Res.* **33**, 21 (2014).
237. Liu, J. et al. The effect of chronic stress on anti-angiogenesis of sunitinib in colorectal cancer models. *Psychoneuroendocrinology* **52**, 130–142 (2015).
238. Hassan, S. et al.  $\beta_2$ -Adrenoreceptor signaling increases therapy resistance in prostate cancer by upregulating MCL1. *Mol. Cancer Res.* **18**, 1839–1848 (2020).
239. Eng, J. W.-L. et al. Housing temperature-induced stress drives therapeutic resistance in murine tumour models through  $\beta_2$ -adrenergic receptor activation. *Nat. Commun.* **6**, 6426 (2015).
- This preclinical study in pancreatic cancer models demonstrates that the ambient housing temperature of laboratory mice can cause chronic adrenergic stress, which in turn can lead to resistance to cytotoxic therapies, but this effect can be reversed by blockade of  $\beta$ -adrenergic signalling. This study supports the potential beneficial effects of  $\beta$ -blockade in the context of cancer therapy.**
240. Chen, M. et al. Adrenergic stress constrains the development of anti-tumor immunity and abscopal responses following local radiation. *Nat. Commun.* **11**, 1821 (2020).
241. Shi, M. et al. Catecholamine-induced  $\beta_2$ -adrenergic receptor activation mediates desensitization of gastric cancer cells to trastuzumab by upregulating MUC4 expression. *J. Immunol.* **190**, 5600–5608 (2013).
242. Liu, D. et al.  $\beta_2$ -AR signaling controls trastuzumab resistance-dependent pathway. *Oncogene* **35**, 47–58 (2016).
243. Zhang, C. et al. Clinical and mechanistic aspects of glucocorticoid-induced chemotherapy resistance in the majority of solid tumors. *Cancer Biol. Ther.* **6**, 278–287 (2007).
244. Arora, V. K. et al. Glucocorticoid receptor confers resistance to antiandrogens by bypassing androgen receptor blockade. *Cell* **155**, 1309–1322 (2013).
245. Skor, M. N. et al. Glucocorticoid receptor antagonism as a novel therapy for triple-negative breast cancer. *Clin. Cancer Res.* **19**, 6163–6172 (2013).
246. Zhang, C. et al. Corticosteroids induce chemotherapy resistance in the majority of tumour cells from bone, brain, breast, cervix, melanoma and neuroblastoma. *Int. J. Oncol.* **29**, 1295–1301 (2006).
- This comprehensive screen identifies glucocorticoid-induced chemotherapy resistance in numerous human carcinoma cell lines.**
247. Fiala, O. et al. Incidental use of  $\beta$ -blockers is associated with outcome of metastatic colorectal cancer patients treated with bevacizumab-based therapy: a single-institution retrospective analysis of 514 patients. *Cancers* **11**, 1856 (2019).
248. Chaudhary, K. R. et al. Effects of  $\beta$ -adrenergic antagonists on chemoradiation therapy for locally advanced non-small cell lung cancer. *J. Clin. Med.* **8**, 575 (2019).
249. Wang, H. et al. Improved survival outcomes with the incidental use of  $\beta$ -blockers among patients with non-small-cell lung cancer treated with definitive radiation therapy. *Ann. Oncol.* **24**, 1312–1319 (2013).
250. Kokolus, K. M. et al.  $\beta$  blocker use correlates with better overall survival in metastatic melanoma patients and improves the efficacy of immunotherapies in mice. *Oncoimmunology* **7**, e1405205 (2018).
251. Navari, R. M. & Aapro, M. Antiemetic prophylaxis for chemotherapy-induced nausea and vomiting. *N. Engl. J. Med.* **374**, 1356–1367 (2016).
252. Pufall, M. A. in *Glucocorticoid Signaling* (eds Wang, J. C. & Harris, C.) 315–333 (Springer, 2015).
253. Boutros, C. et al. Safety profiles of anti-CTLA-4 and anti-PD-1 antibodies alone and in combination. *Nat. Rev. Clin. Oncol.* **13**, 473–486 (2016).
254. Arbour, K. C. et al. Deleterious effect of baseline steroids on efficacy of PD-(L)1 blockade in patients with NSCLC. *J. Clin. Oncol.* **36**, 2872–2878 (2018).
- This retrospective study in patients with non-small-cell lung cancer reports an association between the use of high-dose corticosteroids, reduced efficacy of immune checkpoint inhibitor therapy and poorer clinical outcome, emphasizing the importance of reassessing the prevalent use of synthetic glucocorticoids in patients with cancer.**
255. Scott, S. C. & Pennell, N. A. Early use of systemic corticosteroids in patients with advanced NSCLC treated with nivolumab. *J. Thorac. Oncol.* **13**, 1771–1775 (2018).
256. Fucà, G. et al. Modulation of peripheral blood immune cells by early use of steroids and its association with clinical outcomes in patients with metastatic non-small cell lung cancer treated with immune checkpoint inhibitors. *ESMO Open* **4**, e000457 (2019).
257. Kissane, D. W. et al. Effect of cognitive-existential group therapy on survival in early-stage breast cancer. *J. Clin. Oncol.* **22**, 4255–4260 (2004).
258. Andersen, B. L. et al. Psychologic intervention improves survival for breast cancer patients: a randomized clinical trial. *Cancer* **113**, 3450–3458 (2008).
259. Boesen, E. H. et al. Psychosocial group intervention for patients with primary breast cancer: a randomised trial. *Eur. J. Cancer* **47**, 1363–1372 (2011).
260. Stagl, J. M. et al. A randomized controlled trial of cognitive-behavioral stress management in breast cancer: survival and recurrence at 11-year follow-up. *Breast Cancer Res. Treat.* **154**, 319–328 (2015).
261. Cunningham, A. et al. A randomized controlled trial of the effects of group psychological therapy on survival in women with metastatic breast cancer. *Psychooncology* **7**, 508–517 (1998).
262. Edelman, S., Lemon, J., Bell, D. R. & Kidman, A. D. Effects of group CBT on the survival time of patients with metastatic breast cancer. *Psychooncology* **8**, 474–481 (1999).
263. Kissane, D. W. et al. Supportive-expressive group therapy for women with metastatic breast cancer: survival and psychosocial outcome from a randomized controlled trial. *Psychooncology* **16**, 277–286 (2007).
264. Andersen, B. L. et al. Biobehavioral, immune, and health benefits following recurrence for psychological intervention participants. *Clin. Cancer Res.* **16**, 3270–3278 (2010).
265. Linn, M. W., Linn, B. S. & Harris, R. Effects of counseling for late stage cancer patients. *Cancer* **49**, 1048–1055 (1982).
266. Inlyckij, A., Farber, J., Cheang, M. & Weinerman, B. A randomized controlled trial of psychotherapeutic intervention in cancer patients. *Ann. R. Coll. Physicians Surg. Can.* **27**, 93–96 (1994).
267. Ratcliffe, M. A., Dawson, A. A. & Walker, L. G. Eysenck personality inventory L-scores in patients with Hodgkin's disease and non-Hodgkin's lymphoma. *Psychooncology* **4**, 39–45 (1995).
268. Ross, L. et al. No effect on survival of home psychosocial intervention in a randomized study of Danish colorectal cancer patients. *Psychooncology* **18**, 875–885 (2009).
269. Temel, J. S. et al. Early palliative care for patients with metastatic non-small-cell lung cancer. *N. Engl. J. Med.* **363**, 733–742 (2010).
270. Guo, Z. et al. The benefits of psychosocial interventions for cancer patients undergoing radiotherapy. *Health Qual. Life Outcomes* **11**, 121 (2013).
271. Dhabhar, F. S. Effects of stress on immune function: the good, the bad, and the beautiful. *Immunol. Res.* **58**, 193–210 (2014).
272. Viswanathan, K. & Dhabhar, F. S. Stress-induced enhancement of leukocyte trafficking into sites of surgery or immune activation. *Proc. Natl Acad. Sci. USA* **102**, 5808–5813 (2005).
273. Neeman, E. et al. Stress and skin leukocyte trafficking as a dual-stage process. *Brain Behav. Immun.* **26**, 267–276 (2012).
274. Russell, G. & Lightman, S. The human stress response. *Nat. Rev. Endocrinol.* **15**, 525–534 (2019).
275. Cruz-Topete, D. & Cidlowski, J. A. One hormone, two actions: anti- and pro-inflammatory effects of glucocorticoids. *Neuroimmunomodulation* **22**, 20–32 (2015).
276. Shaashua, L. et al. In vivo suppression of plasma IL-12 levels by acute and chronic stress paradigms: potential mediating mechanisms and sex differences. *Brain Behav. Immun.* **26**, 996–1005 (2012).
277. Baum, A., O'Keeffe, M. K. & Davidson, L. M. Acute stressors and chronic response: the case of traumatic stress. *J. Appl. Soc. Psychol.* **20**, 1643–1654 (1990).
278. Hawley, J. A., Hargreaves, M., Joyner, M. J. & Zierath, J. R. Integrative biology of exercise. *Cell* **159**, 738–749 (2014).
279. Neuffer, P. D. et al. Understanding the cellular and molecular mechanisms of physical activity-induced health benefits. *Cell Metab.* **22**, 4–11 (2015).
280. Brownley, K. A. et al. Sympathoadrenergic mechanisms in reduced hemodynamic stress responses after exercise. *Med. Sci. Sports Exerc.* **35**, 978–986 (2003).
281. Traustadóttir, T., Bosch, P. R. & Matt, K. S. The HPA axis response to stress in women: effects of aging and fitness. *Psychoneuroendocrinology* **30**, 392–402 (2005).
282. Petersen, A. M. W. & Pedersen, B. K. The anti-inflammatory effect of exercise. *J. Appl. Physiol.* **98**, 1154–1162 (2005).
283. Speck, R. M., Courneya, K. S., Mäse, L. C., Duval, S. & Schmitz, K. H. An update of controlled physical activity trials in cancer survivors: a systematic review and meta-analysis. *J. Cancer Surviv.* **4**, 87–100 (2010).
284. Rogers, L. O. et al. Effects of a multicomponent physical activity behavior change intervention on fatigue, anxiety, and depressive symptomatology in breast cancer survivors: randomized trial. *Psychooncology* **26**, 1901–1906 (2017).
285. Mehnert, A. et al. Effects of a physical exercise rehabilitation group program on anxiety, depression, body image, and health-related quality of life among breast cancer patients. *Oncol. Res. Treat.* **34**, 248–253 (2011).
286. Dimeo, F. C., Stieglitz, R. D., Novelli-Fischer, U., Fetscher, S. & Keul, J. Effects of physical activity on the fatigue and psychologic status of cancer patients during chemotherapy. *Cancer* **85**, 2273–2277 (1999).
287. McNeely, M. L. et al. Effects of exercise on breast cancer patients and survivors: a systematic review and meta-analysis. *CMAJ* **175**, 34–41 (2006).
288. Kruijsen-Jaarsma, M., Révész, D., Bierings, M. B., Buffart, L. M. & Takken, T. Effects of exercise on immune function in patients with cancer: a systematic review. *Exec. Immunol. Rev.* **19**, 120–143 (2013).
289. Davies, N., Bateup, L. & Thomas, R. The role of diet and physical activity in breast, colorectal, and prostate cancer survivorship: a review of the literature. *Br. J. Cancer* **105**, S52–S73 (2011).
290. Stout, N. L., Baima, J., Swisher, A. K., Winters-Stone, K. M. & Welsh, J. A systematic review of exercise systematic reviews in the cancer literature (2005–2017). *PMR* **9**, S347–S384 (2017).
291. Hanns, P., Paczulla, A. M., Medinger, M., Konantz, M. & Lengerke, C. Stress and catecholamines modulate the bone marrow microenvironment to promote tumorigenesis. *Cell Stress* **3**, 221 (2019).
292. Dethlefsen, C. et al. Exercise-induced catecholamines activate the hippo tumor suppressor pathway to

- reduce risks of breast cancer development. *Cancer Res.* **77**, 4894–4904 (2017).
293. Pedersen, L. et al. Voluntary running suppresses tumor growth through epinephrine- and IL-6-dependent NK cell mobilization and redistribution. *Cell Metab.* **23**, 554–562 (2016).
  294. Song, Y. et al. Enriching the housing environment for mice enhances their NK cell antitumor immunity via sympathetic nerve-dependent regulation of NKG2D and CCR5. *Cancer Res.* **77**, 1611–1622 (2017).
  295. Graff, R. M. et al.  $\beta_2$ -Adrenergic receptor signaling mediates the preferential mobilization of differentiated subsets of CD8<sup>+</sup> T-cells, NK-cells and non-classical monocytes in response to acute exercise in humans. *Brain Behav. Immun.* **74**, 143–153 (2018).
  296. Devalon, M. et al. DOPA in plasma increases during acute exercise and after exercise training. *J. Lab. Clin. Med.* **114**, 321–327 (1989).
  297. Yamaguchi, K., Takagi, Y., Aoki, S., Futamura, M. & Saji, S. Significant detection of circulating cancer cells in the blood by reverse transcriptase-polymerase chain reaction during colorectal cancer resection. *Ann. Surg.* **232**, 58–65 (2000).
  298. Hashimoto, M. et al. Significant increase in circulating tumour cells in pulmonary venous blood during surgical manipulation in patients with primary lung cancer. *Interact. Cardiovasc. Thorac. Surg.* **18**, 775–783 (2014).
  299. O'Reilly, M. S. et al. Endostatin: an endogenous inhibitor of angiogenesis and tumor growth. *Cell* **88**, 277–285 (1997).
  300. O'Reilly, M. S. et al. Angiostatin: a novel angiogenesis inhibitor that mediates the suppression of metastases by a Lewis lung carcinoma. *Cell* **79**, 315–328 (1994).
  301. Abramovitch, R., Marikovsky, M., Meir, G. & Neeman, M. Stimulation of tumour growth by wound-derived growth factors. *Br. J. Cancer* **79**, 1392–1398 (1999).
  302. Pascual, M. et al. Randomized clinical trial comparing inflammatory and angiogenic response after open versus laparoscopic curative resection for colonic cancer. *Br. J. Surg.* **98**, 50–59 (2011).
  303. Garssen, B., Boomsma, M. F. & Beelen, R. H. Psychological factors in immunomodulation induced by cancer surgery: a review. *Biol. Psychol.* **85**, 1–13 (2010).
  304. Cata, J. P. et al. Intraoperative use of dexmedetomidine is associated with decreased overall survival after lung cancer surgery. *J. Anaesthesiol. Clin. Pharmacol.* **33**, 317 (2017).
  305. Lavon, H. et al. Dexmedetomidine promotes metastasis in rodent models of breast, lung, and colon cancers. *Br. J. Anaesth.* **120**, 188–196 (2018).
  306. Del Mastro, L. et al. Impact of two different dose-intensity chemotherapy regimens on psychological distress in early breast cancer patients. *Eur. J. Cancer* **38**, 359–366 (2002).
  307. Vyas, D., Laput, G. & Vyas, A. K. Chemotherapy-enhanced inflammation may lead to the failure of therapy and metastasis. *Onco. Targets Ther.* **7**, 1015 (2014).
  308. Shaked, Y. Balancing efficacy of and host immune responses to cancer therapy: the yin and yang effects. *Nat. Rev. Clin. Oncol.* **13**, 611 (2016).
  309. Antoni, M. H. et al. How stress management improves quality of life after treatment for breast cancer. *J. Consul. Clin. Psychol.* **74**, 1143 (2006).
  310. Andersen, B. L. et al. Distress reduction from a psychological intervention contributes to improved health for cancer patients. *Brain Behav. Immun.* **21**, 953–961 (2007).
  311. Riba, M. B. et al. Distress management, version 3.2019, NCCN clinical practice guidelines in oncology. *J. Natl Compr. Cancer Netw.* **17**, 1229–1249 (2019).
  312. Buffart, L. M. et al. Physical and psychosocial benefits of yoga in cancer patients and survivors, a systematic review and meta-analysis of randomized controlled trials. *BMC Cancer* **12**, 1–21 (2012).
  313. Bower, J. E. et al. Yoga reduces inflammatory signaling in fatigued breast cancer survivors: a randomized controlled trial. *Psychoneuroendocrinology* **43**, 20–29 (2014).
  314. Witek-Janusek, L. et al. Effect of mindfulness based stress reduction on immune function, quality of life and coping in women newly diagnosed with early stage breast cancer. *Brain Behav. Immun.* **22**, 969–981 (2008).
  315. Bower, J. E. et al. Mindfulness meditation for younger breast cancer survivors: a randomized controlled trial. *Cancer* **121**, 1231–1240 (2015).
  316. Antoni, M. H. *Stress Management Intervention for Women with Breast Cancer* (American Psychological Association, 2003).
  317. Antoni, M. H. et al. Cognitive behavioral stress management effects on psychosocial and physiological adaptation in women undergoing treatment for breast cancer. *Brain Behav. Immun.* **23**, 580–591 (2009).

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